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**Pr Boualem Sendid**

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**DU d'Antibiothérapie et Chimiothérapie Anti-Infectieuse, 2023-2024**

# Epidemiology of Invasive Fungal Infections

Open Forum Infectious Diseases

MAJOR ARTICLE

## Epidemiology and Clinical Features of Invasive Fungal Infection in a US Health Care Network

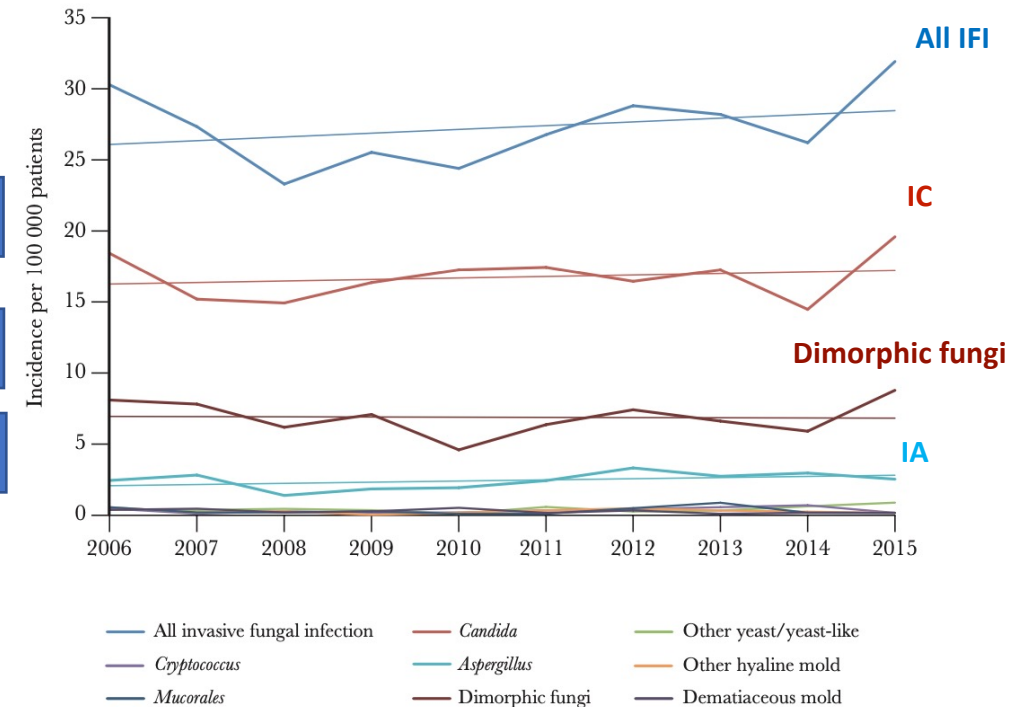
Brandon J. Webb,<sup>1,2</sup> Jeffrey P. Ferraro,<sup>3,4</sup> Susan Rea,<sup>3</sup> Stephanie Kaufusi,<sup>3,5</sup> Bruce E. Goodman,<sup>3,5</sup> and James Spalding<sup>6</sup>



Total of 3374 IFI episodes occurred in 3154 patients. The mean incidence of IFI was 27.2 cases/100 000 patients per year.

Table 1. Incidence (per 100 000 Patients) of IFI by Category and Year




















	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2006–2015
All IFIs	30.2	27.2	23.2	25.4	24.3	26.7	28.7	28.1	26.1	31.8	27.2
<i>Candida</i>	18.3	15.1	14.9	16.3	17.2	17.4	16.4	17.2	14.4	19.5	15.0
Other yeast/yeast-like	0.4	0.3	0.4	0.3	0.1	0.3	0.2	0.3	0.3	0.3	0.4
<i>Cryptococcus</i>	0.4	0.1	0.3	0.2	0.2	0.3	0.4	0.5	0.7	0.1	0.3
<i>Aspergillus</i>	2.4	2.8	1.4	1.8	1.9	2.4	3.3	2.7	2.9	2.5	2.4
Other hyaline mold	0.4	0.2	0.3	0.0	0.2	0.3	0.5	0.3	0.2	0.1	0.2
<i>Mucorales</i>	0.5	0.2	0.2	0.2	0.1	0.1	0.5	0.9	0.2	0.1	0.3
Dimorphic fungi	8.1	7.8	6.1	7.0	4.6	6.3	7.4	6.6	5.9	8.7	6.9
Dematiaceous mold	0.3	0.4	0.2	0.3	0.5	0	0.3	0.1	0.2	0.1	0.2



Webb BJ et al. Open Forum Infect Dis. 2018

Figure 1. Incidence of invasive fungal infection (IFI) by category (per 100 000 patients).

# WHO fungal priority pathogens list to guide research, development and public health action

Critical group	High group	Medium group
 <i>Cryptococcus neoformans</i>	 <i>Nakaseomyces glabrata</i> ( <i>Candida glabrata</i> )	 <i>Scedosporium</i> spp.
 <i>Candida auris</i>	 <i>Histoplasma</i> spp.	 <i>Lomentospora prolificans</i>
 <i>Aspergillus fumigatus</i>	 Eumycetoma causative agents	 <i>Coccidioides</i> spp.
 <i>Candida albicans</i>	 Mucorales	 <i>Pichia kudriavzevii</i> ( <i>Candida krusei</i> )
	 <i>Fusarium</i> spp.	 <i>Cryptococcus gattii</i>
	 <i>Candida tropicalis</i>	 <i>Talaromyces marneffei</i>
	 <i>Candida parapsilosis</i>	 <i>Pneumocystis jirovecii</i>
		 <i>Paracoccidioides</i> spp.

## TUBERCULOSIS: A GLOBAL PRIORITY FOR RESEARCH AND DEVELOPMENT

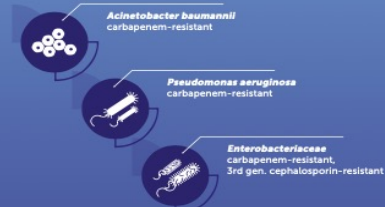
### FIVE REASONS WHY

- 1** Tuberculosis (TB) is the number one global infectious disease killer today, causing 1.8 million deaths per year. Drug-resistant TB is the most common and lethal airborne AMR disease worldwide today, responsible for 250,000 deaths each year.
- 2** Patients with multidrug-resistant TB (MDR-TB) need complex and prolonged multidrug treatment with costly, highly toxic, and much less effective second-line medicines. There is a limited number of second-line medicines to treat MDR-TB and only 52% of patients are successfully treated globally.
- 3** In about 50% of MDR-TB patients worldwide, treatment regimens are already compromised by second-line drug resistance. Treatment of extensively drug-resistant disease (XDR-TB) is successful in only one in three patients at best.
- 4** Patients with M/XDR-TB face agonising, prolonged suffering and often permanent disability while on treatment, compounded by devastating economic hardship, stigma and discrimination.
- 5** Only two new antibiotics for treatment of MDR-TB have reached the market in over 70 years. R&D investment in TB – seriously underfunded – is at its lowest level since 2008.

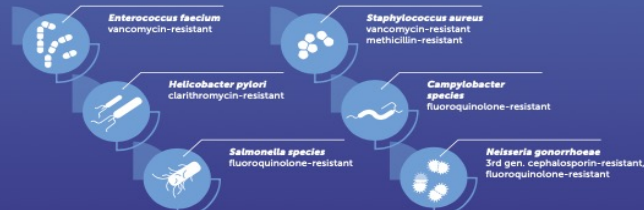
<sup>1</sup> MDR-TB – multidrug-resistant tuberculosis, that does not respond to at least isoniazid and rifampicin, the two most powerful first-line anti-TB medicines.  
<sup>2</sup> XDR-TB – extensively drug-resistant tuberculosis, defined as MDR-TB plus resistance to fluoroquinolones and injectable second-line anti-TB medicines.

### OTHER PRIORITY PATHOGENS

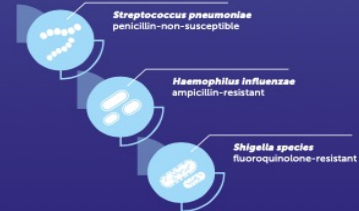
#### CRITICAL PRIORITY



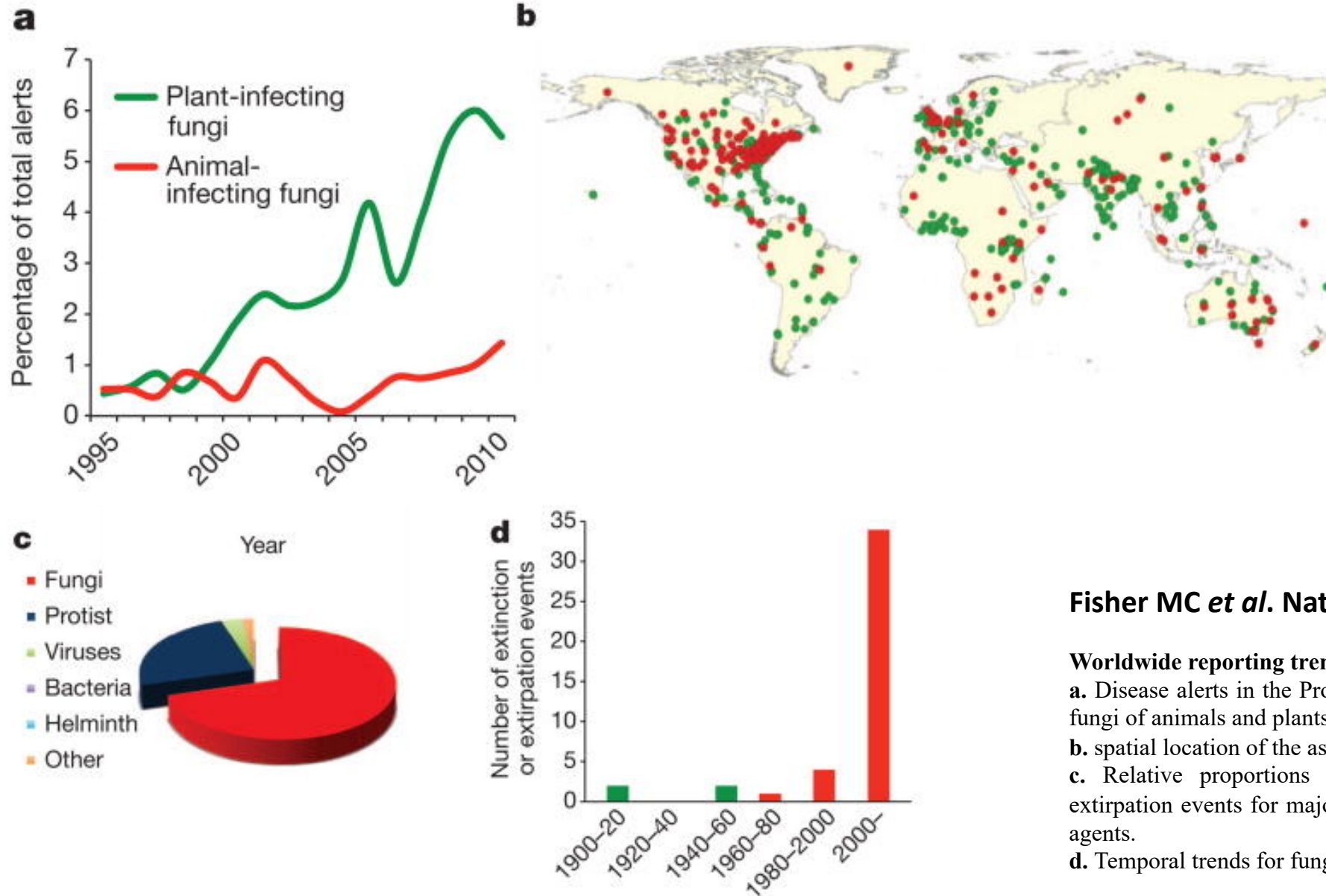
#### HIGH PRIORITY



#### MEDIUM PRIORITY



# Emerging Fungal Threats to Animal, Plant and Ecosystem Health



Fisher MC *et al.* Nature. 2012

## Worldwide reporting trends in fungal EIDs.

- a. Disease alerts in the ProMED database for pathogenic fungi of animals and plants, and the
- b. spatial location of the associated reports.
- c. Relative proportions of species extinction and/or extirpation events for major classes of infectious disease agents.
- d. Temporal trends for fungal pathogens

# Mycoses superficielles



Candidose buccale



Candidose génitale



Candidose interdigito-palmaire



Pityriasis versicolor



Pityriasis versicolor

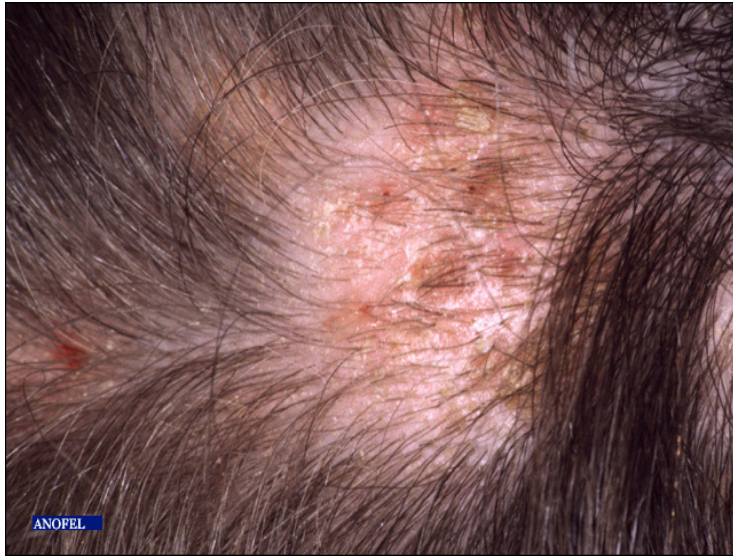


Herpes circiné

# Mycoses superficielles

## Atteintes à dermatophytes

*Teignes microsporiques*



*Teignes trichophytiques*



*Favus*



*Teignes inflammatoires: kérion*



*Teignes inflammatoires: sycosis*



# Diagnostic des mycoses superficielles

Diagnostic aisé fondé sur les méthodes conventionnelles :

- Prélèvements (écouvillons, squames, cheveux, ongles...)
- Examen direct (éléments morphologiques d'orientation)
- Mise en culture (isolement de l'agent pathogène)
- Identification (Méthodes phénotypiques, MALDI-TOF)

# Examen direct à l'état frais ou après coloration

Urines



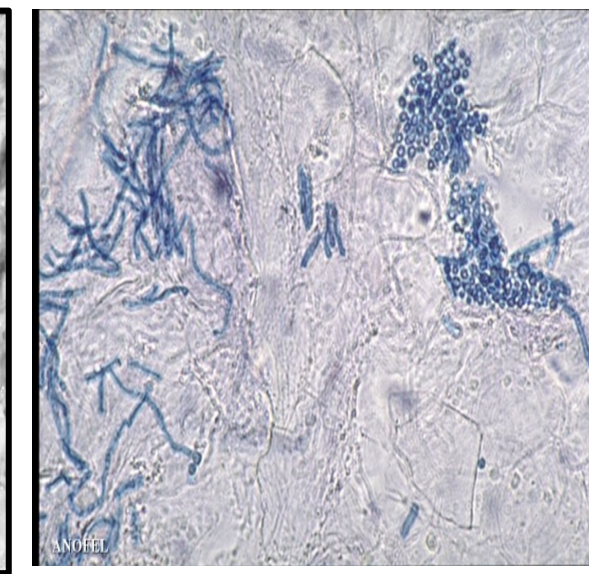
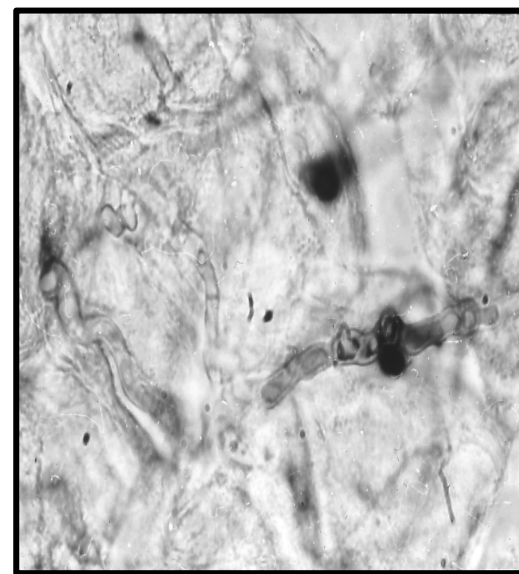
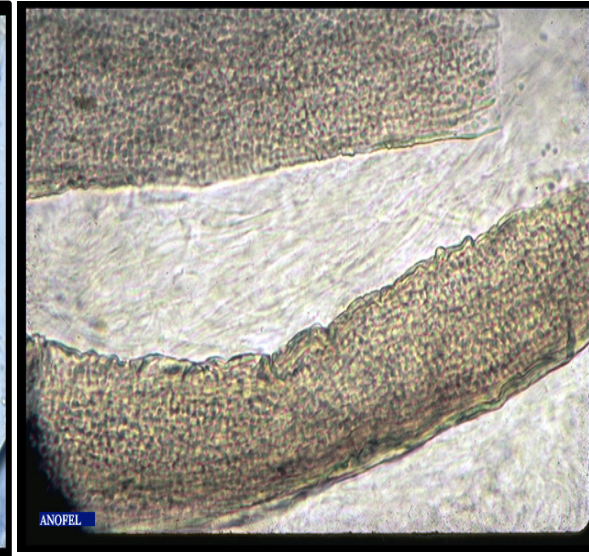
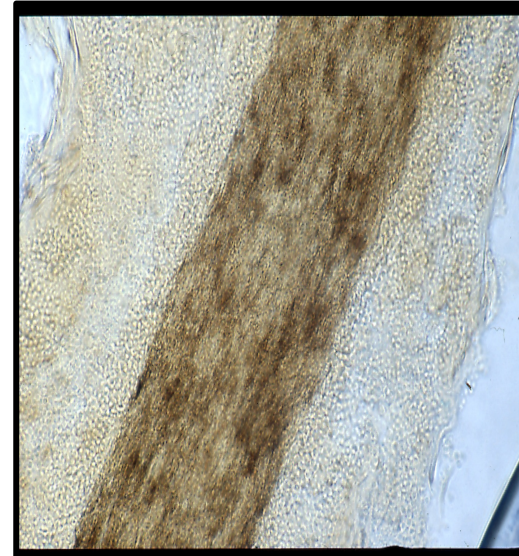
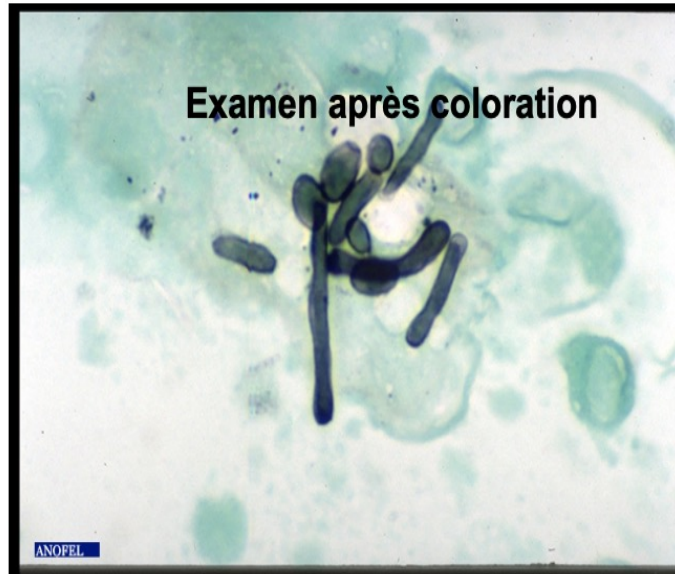
Squames



Selles



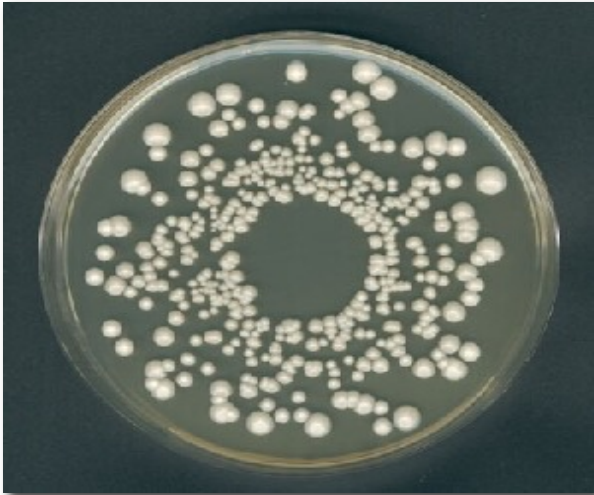
Examen après coloration





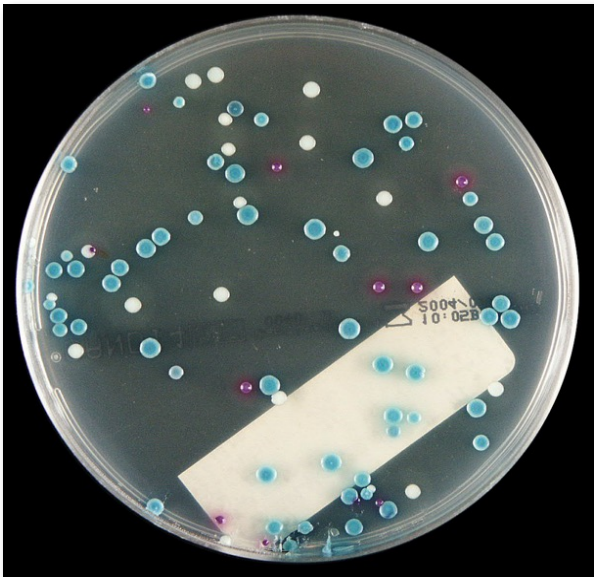
# Identification présomptive : culture sur milieu chromogéniques

## Milieu Sabouraud

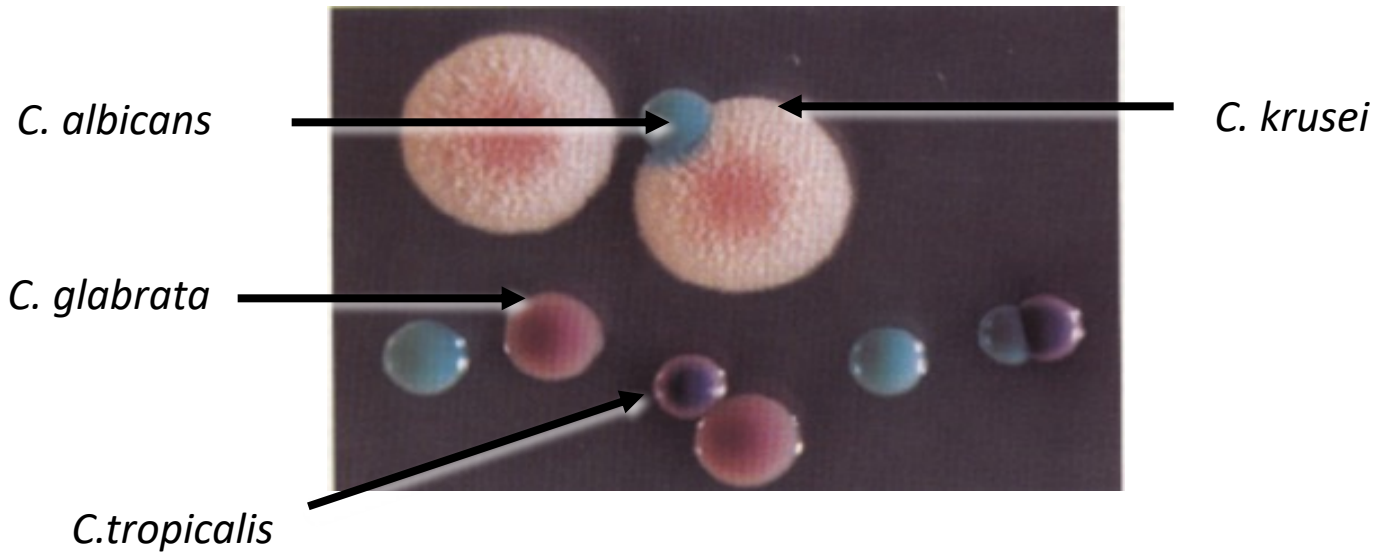


## Avantages

- Isolement et identification des 4 principales espèces
- Détection des mélanges d'espèces
- Différents milieux : CHROMagar, CandiSelect4  
Candichrom, albicans ID, fluoroplate...



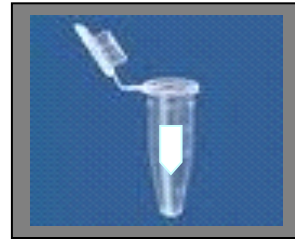
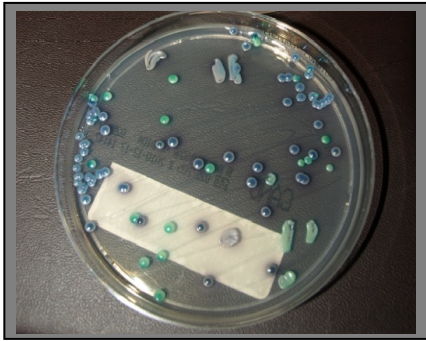
Milieu CandiSelect4®



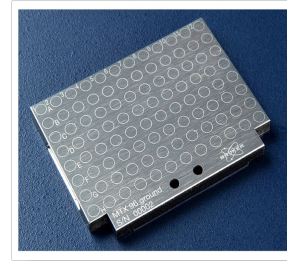
Milieu CHROMagar™ Candida

# Identification par spectrométrie de masse

Extraction acide formique  
Acétonitrile



Extrait + matrice



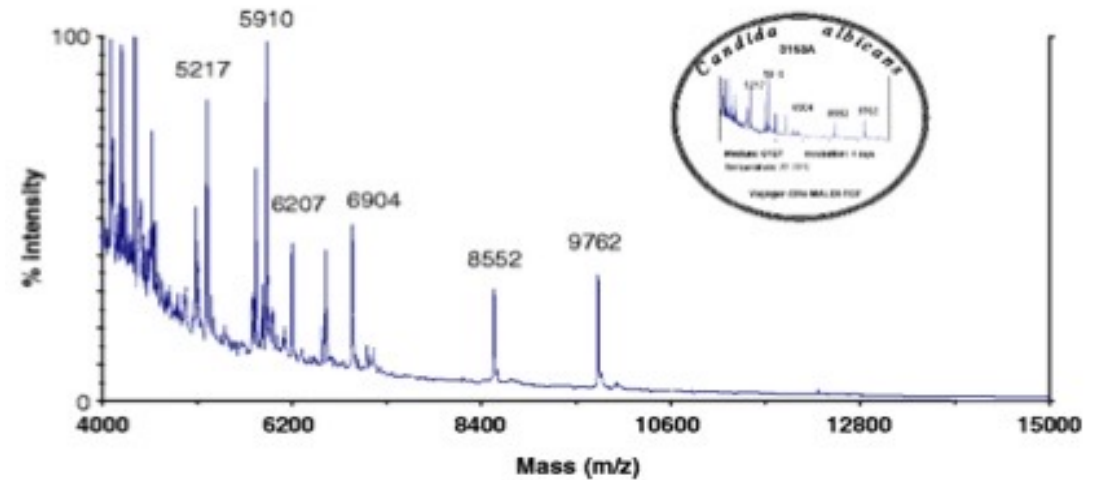
Analyse



15 min

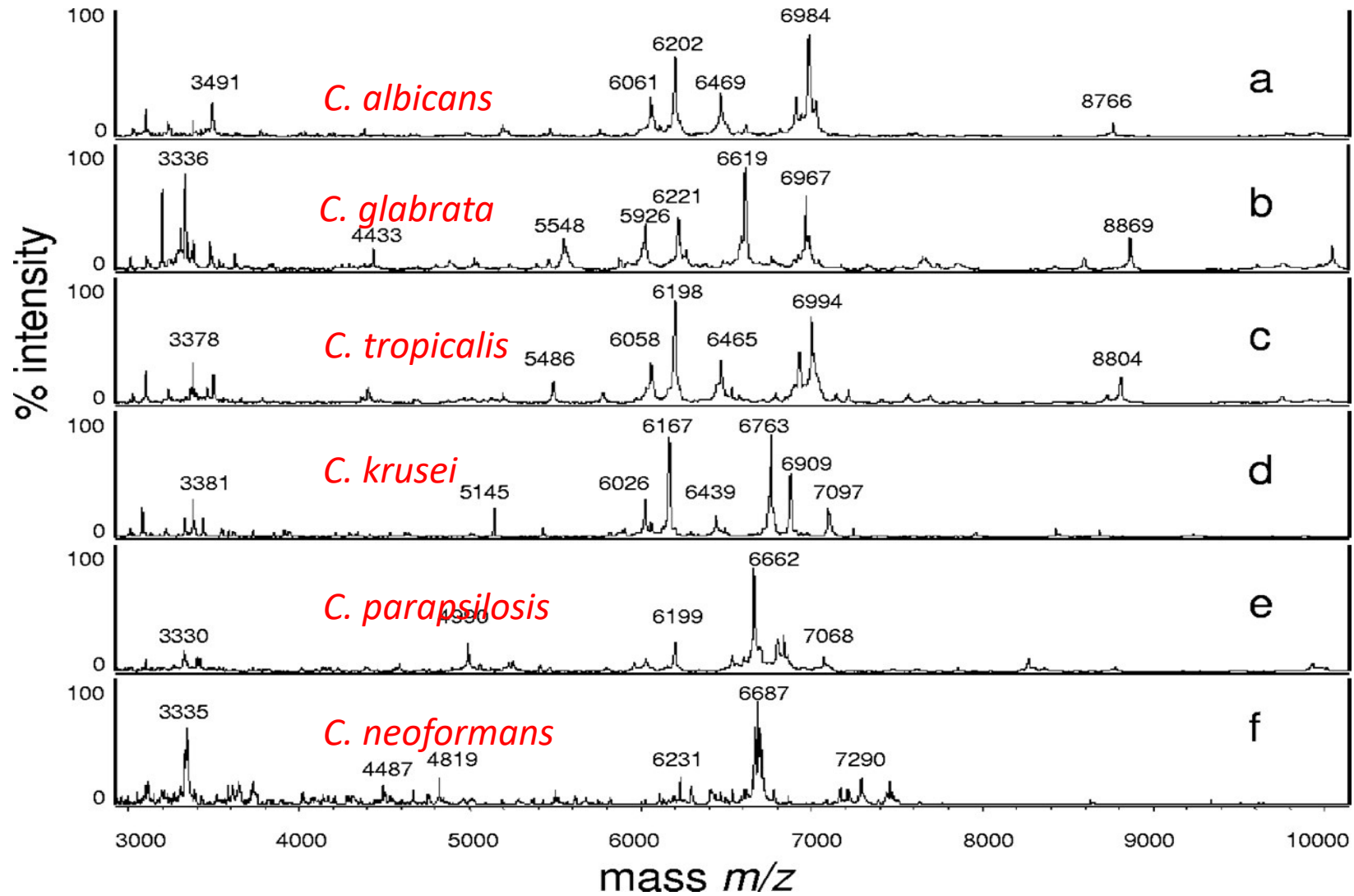


MALDI-TOF BiFlex III®  
(Bruker Daltonics™)



Profil peptidique spécifique d'espèce

Identification par spectrométrie de masse (MALDI-TOF: Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry)

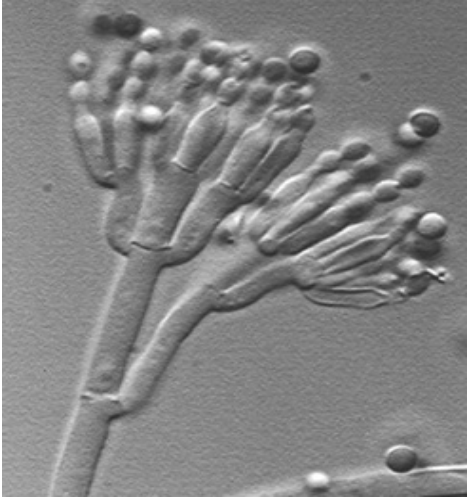


# Examen microscopique des cultures

*Alternaria*



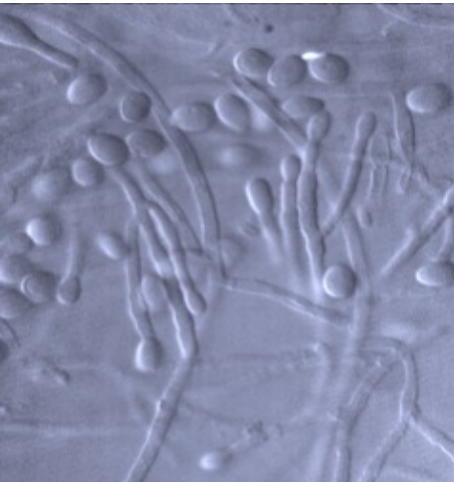
*Penicillium*



*Fusarium*



*Scedosporium*



*Aspergillus*



*Trichophyton*



# **Diagnostic biologique des infections fongiques Invasives**

## Statistics of the 10 most significant human invasive fungal infections

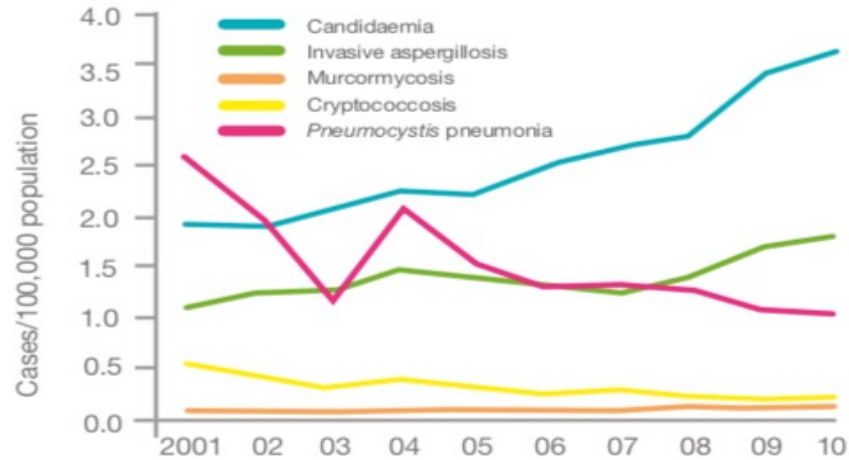
Disease (most common species)	Location	Estimated life-threatening infections/ year at that location*	Mortality rates (% in infected populations)*
Opportunistic invasive mycoses			
Aspergillosis ( <i>A. fumigatus</i> )	worldwide	>200,000	30-95
Candidiasis ( <i>C. albicans</i> )	worldwide	>400,000	46-75
Cryptococcosis ( <i>Cryptococcus neoformans</i> )	worldwide	>1,400,000	20-70
Mucormycosis ( <i>Rhizopus oryzae</i> )	worldwide	>10,000	30-90
Pneumocystis ( <i>P. jirovecii</i> )	worldwide	>400,000	20-80

**IFI kill 1,5 million /year  
= Tuberculosis, more than malaria**

# Impact médico-économiques des Infections Fongiques Invasives

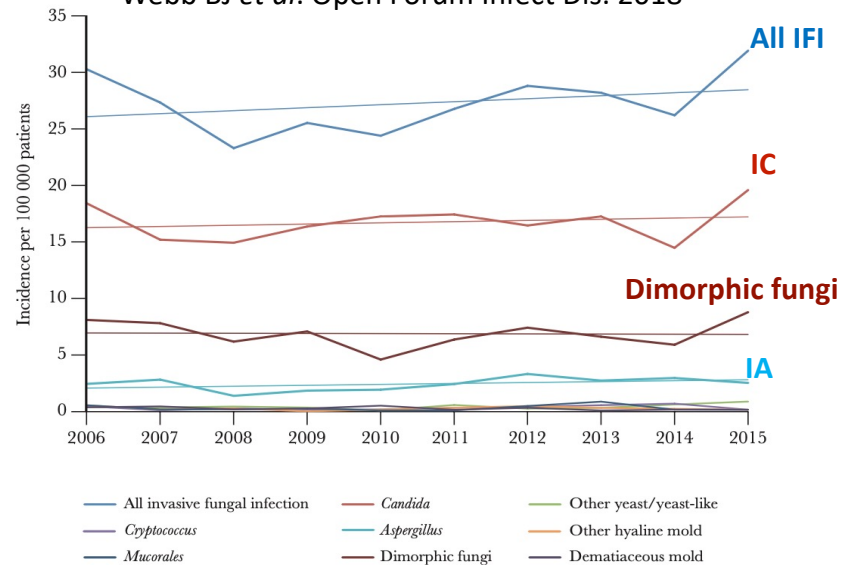
Rates of invasive fungal infections in France (2001-2010)

Bitar D et al., Bull Epidemiol Hebdo, 2013



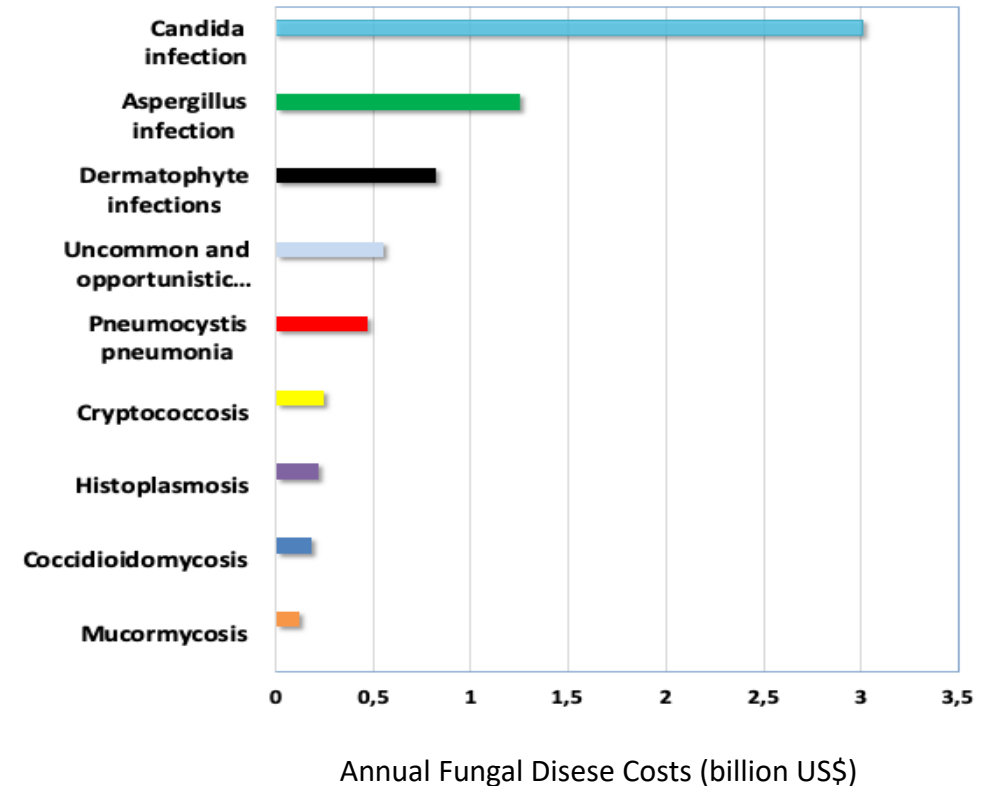
Rates of invasive fungal infections in USA (2006-2015)

Webb BJ et al. Open Forum Infect Dis. 2018

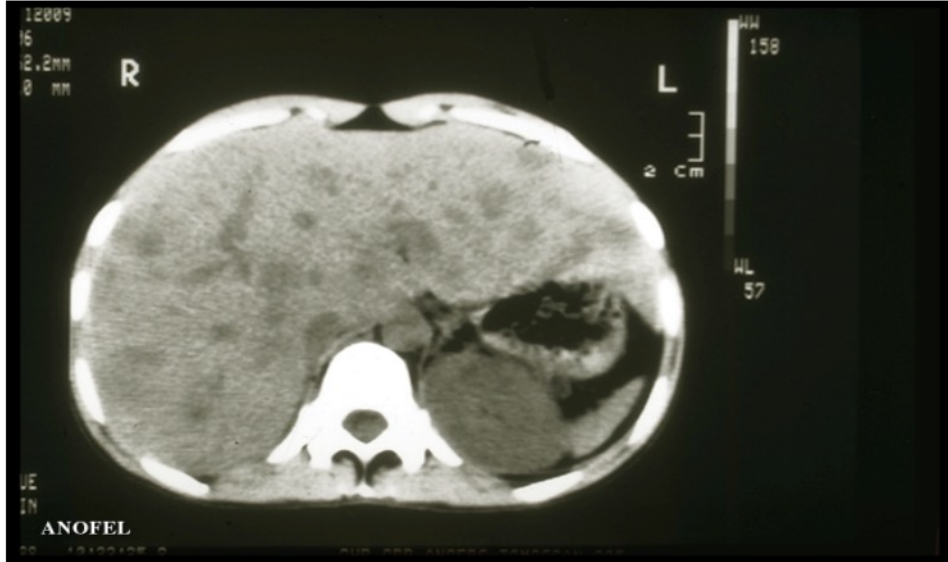


Annual Estimated Total Inpatient and Outpatient Costs for Fungal Diseases in United States\*

\*Kaitlin B et al. Clin Infect Dis 2020



# Candidoses invasives



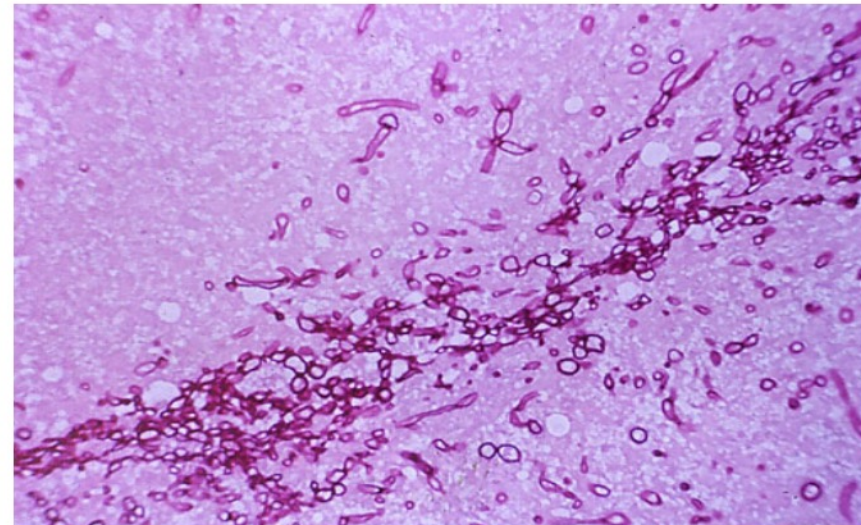
**Candidose hépatique : Importance de l'imagerie**



**Examen macroscopique**



**Micro-abcès**

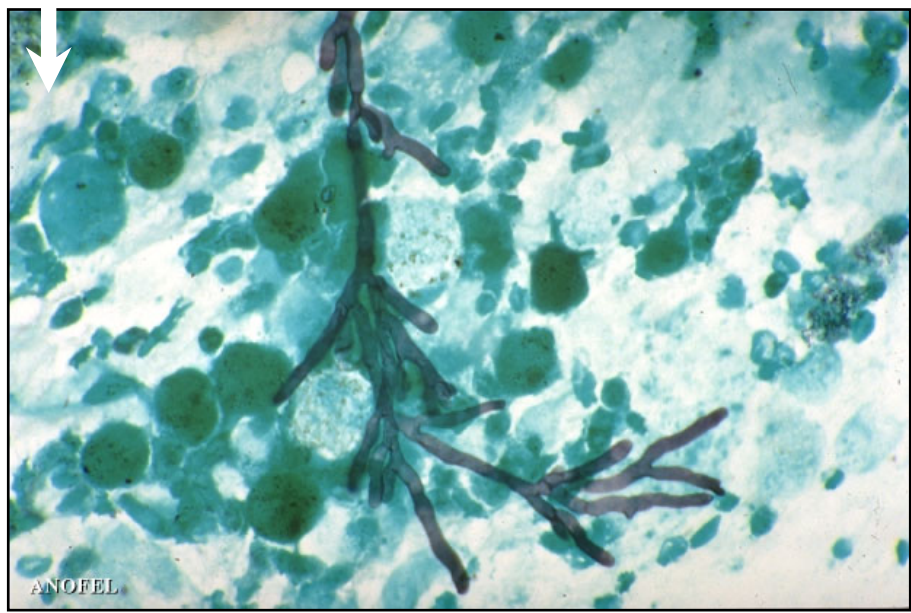
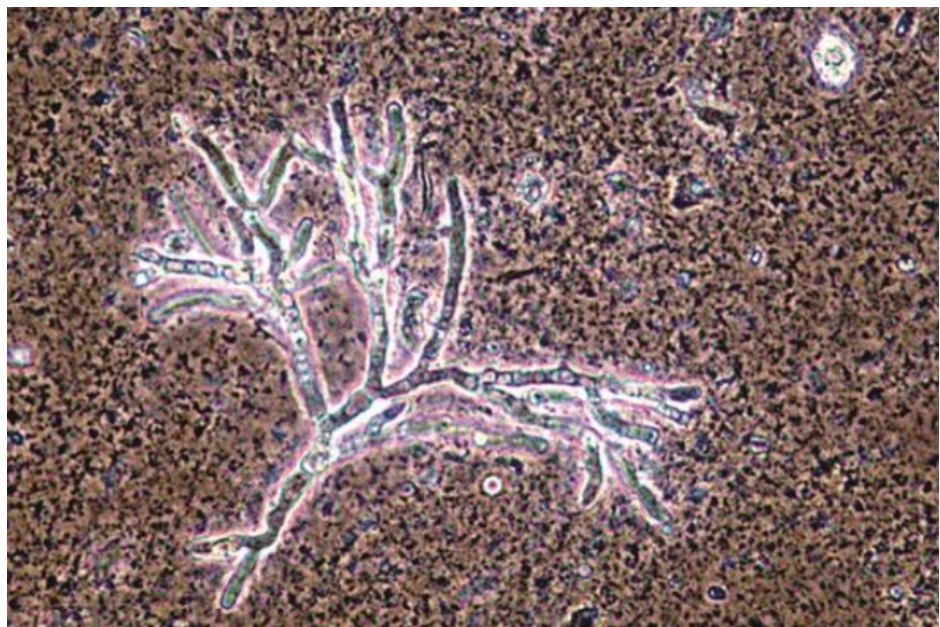
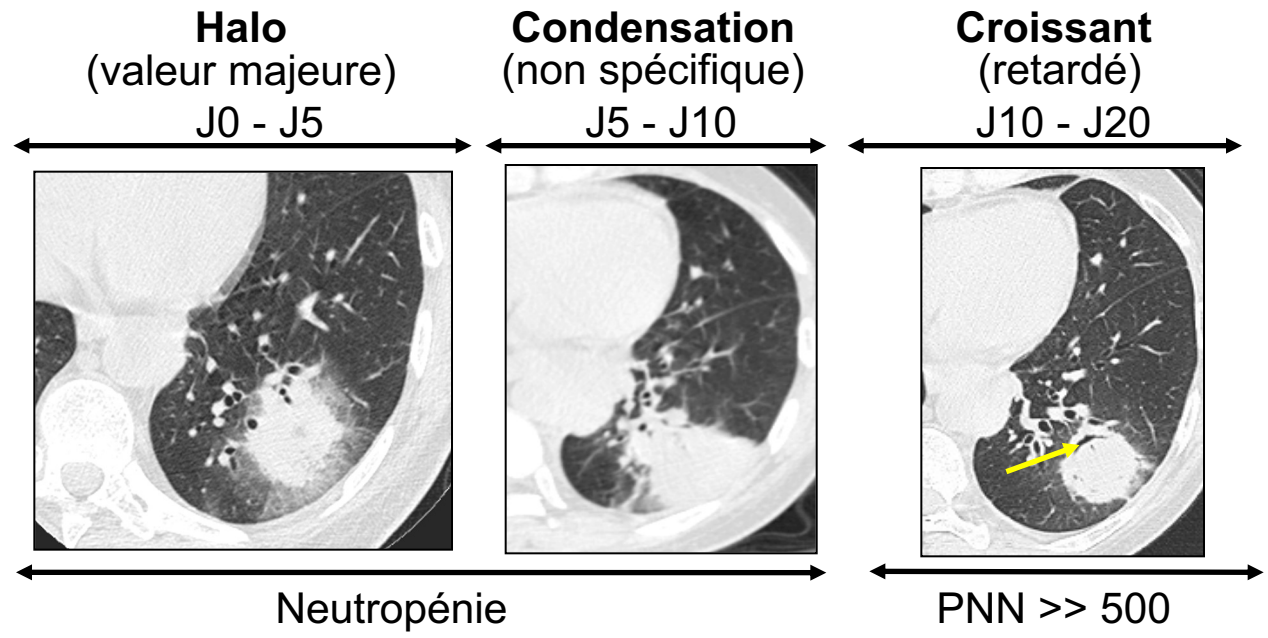


**Examen histologique : Filaments + levures**



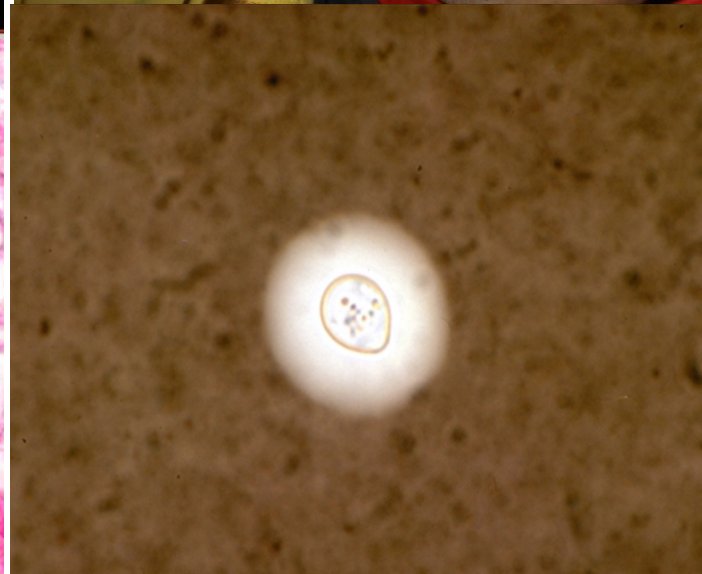
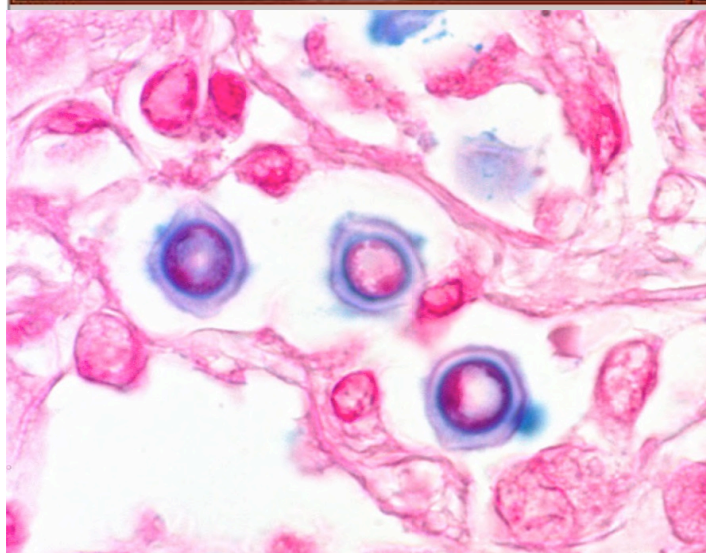
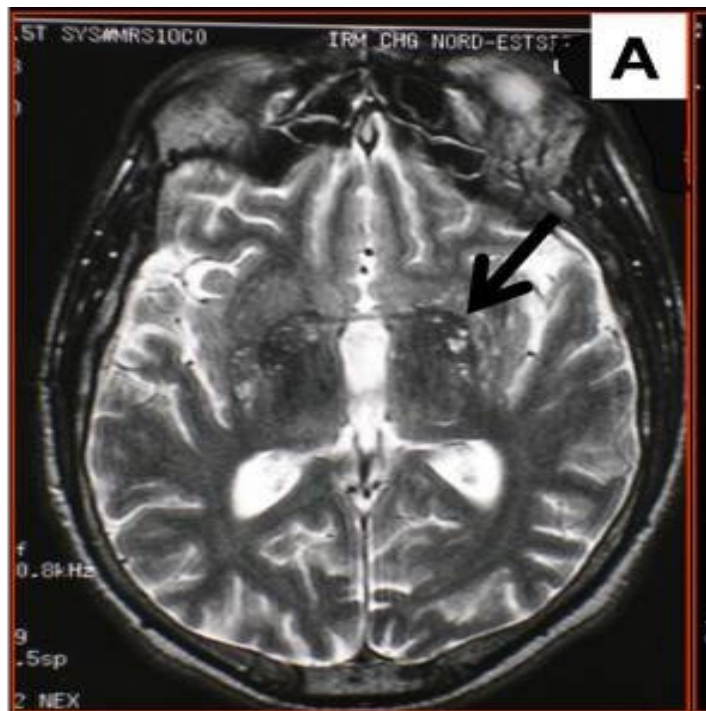
# Mycoses profondes

Aspergillose invasive



Examen direct du LBA

# Cryptococcose neuroméningée



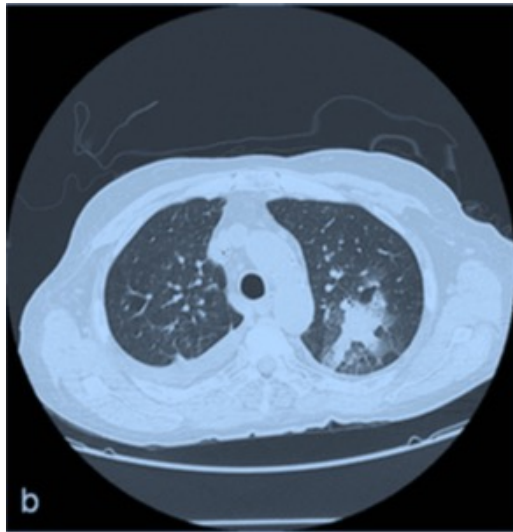
# Mucormycoses invasives



Mucormycose rhino-orbito-cérébrales



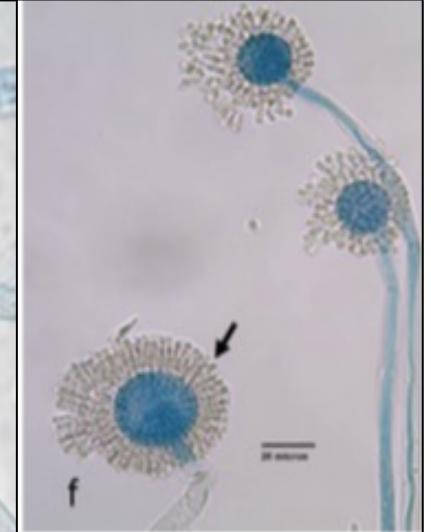
Mucormycose post-traumatique



b



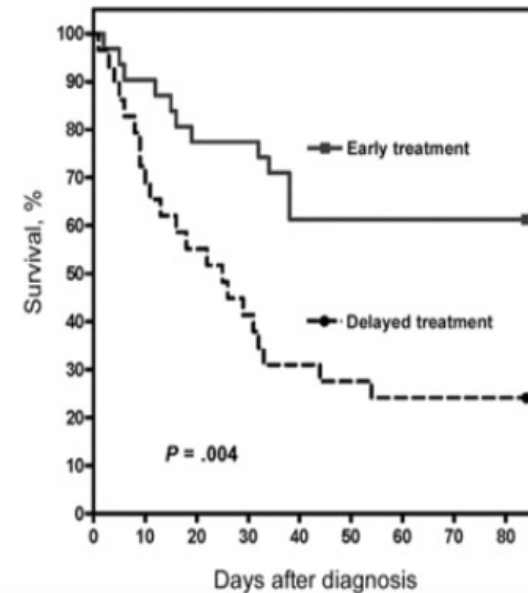
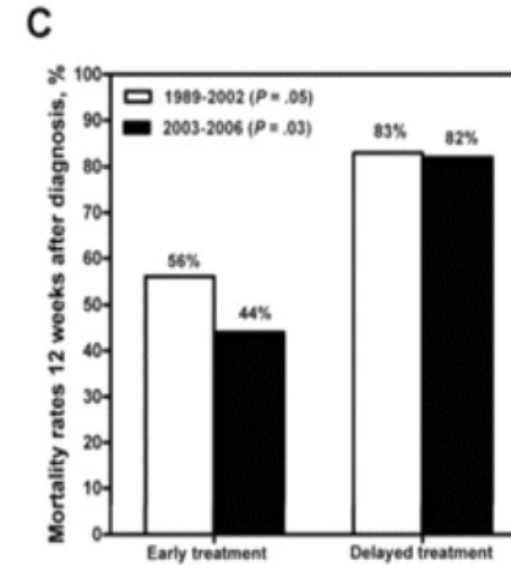
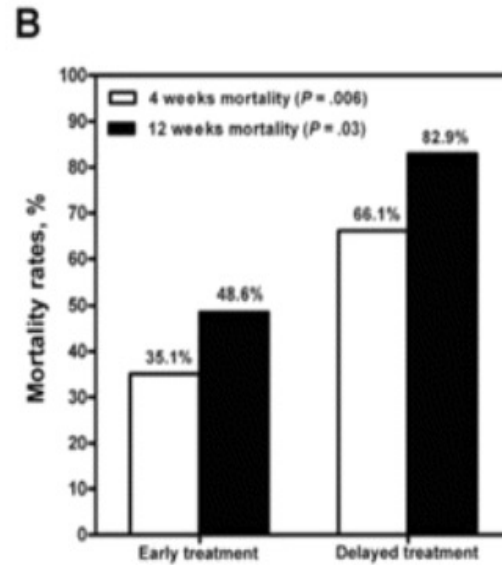
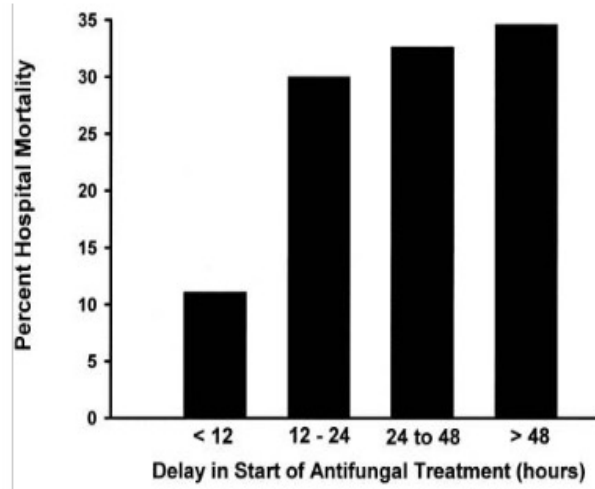
d



f

Mucormycose disséminée

# Importance du diagnostic précoce des infections fongiques invasives



Morrell *et al.* 2005. AAC; Greene *et al.* CID. 2008; Chamilos *et al.* CID. 2008

# Approches et outils disponibles

Mycologie  
conventionnelle

Histologie

Alternatives à la culture (biologie moléculaire,  
Marqueurs sériques)

Examen  
direct

Culture

Séquençage  
panfongique

Détection  
d'ADN  
circulant ou  
*in situ* (RT-  
PCR)

Détection des  
antigène  
circulant/  
Anticorps  
spécifiques

- Orientation  
diagnostique  
- Peu sensible

- Valeur  
diagnostique  
: sites stériles  
- Tests de  
sensibilité  
aux ATF

- Spécifique (Ac  
monoclonaux),  
- Invasif  
- Diagnostic de  
certitude

- Sensibilité  
faible

Standardisation ?  
Résultat  
dépendant de  
matrice, gène  
cible, type de PCR

Performances  
variables selon les  
population de  
patients

## Diagnostic biologique des candidoses invasives

# CONVENTIONAL METHODS

## Culture

- Blood
- other sterile body sites
- urines
- other sites (Diff. diagnosis colonisation- infection)

Insensitive (less than 50% for BC), time consuming for antifungal susceptibility testing

Time for species identification reduced by MALDI-TOF

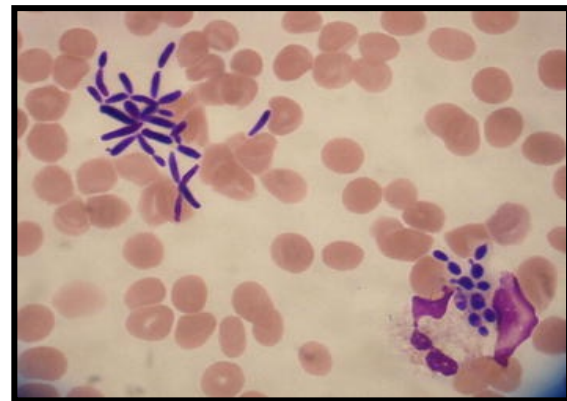
Sampling (10-20 mL of Blood)



Incubator (35°C, > 5 days)

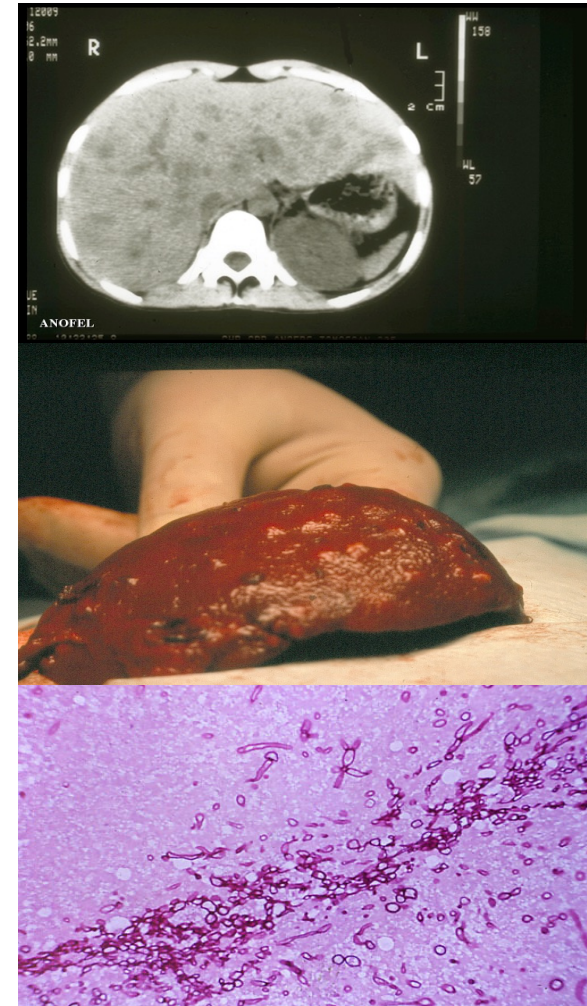


Microscopic examination



# HISTOLOGY

- biopsies (liver, skin)
- low yield (patients under AFT)
- rarely performed in patients with aplasia





# NON-CULTURE BASED METHODS

## ***Candida* – indirect tests**

- antigen
- antibodies
- fungal DNA detection

**Mostly evaluated in onco-hematology and ICU patients**

**Antigen and PCR tests evaluated in neonates**

**Very few commercially available tests**

# Comparison of (1→3)-β-D-Glucan, Mannan/Anti-Mannan Antibodies, and Cand-Tec *Candida* Antigen as Serum Biomarkers for Candidemia

56 candidemia, 200 controls (100 bacteremia, 100 sterile blood cultures)

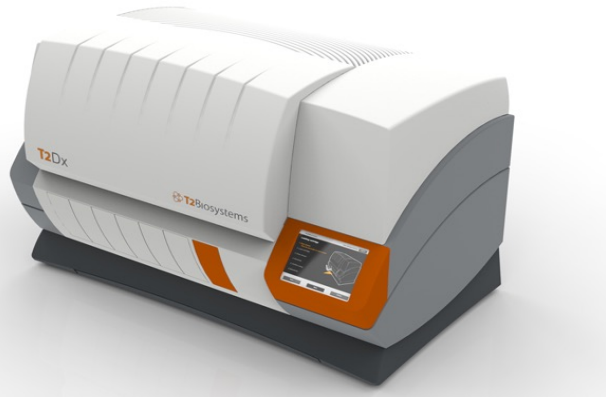
	Sensitivity, %	Specificity, %
BDG	87.5	85.5
Mn	58.9	97.5
Anti-Mn	62.5	65
Mn-Anti-Mn	89.3	63
BDG+Mn	89.3	85
Cand Tec <i>Candida</i> Ag	13	93.9

Held J, J Clin Microbiol 2013

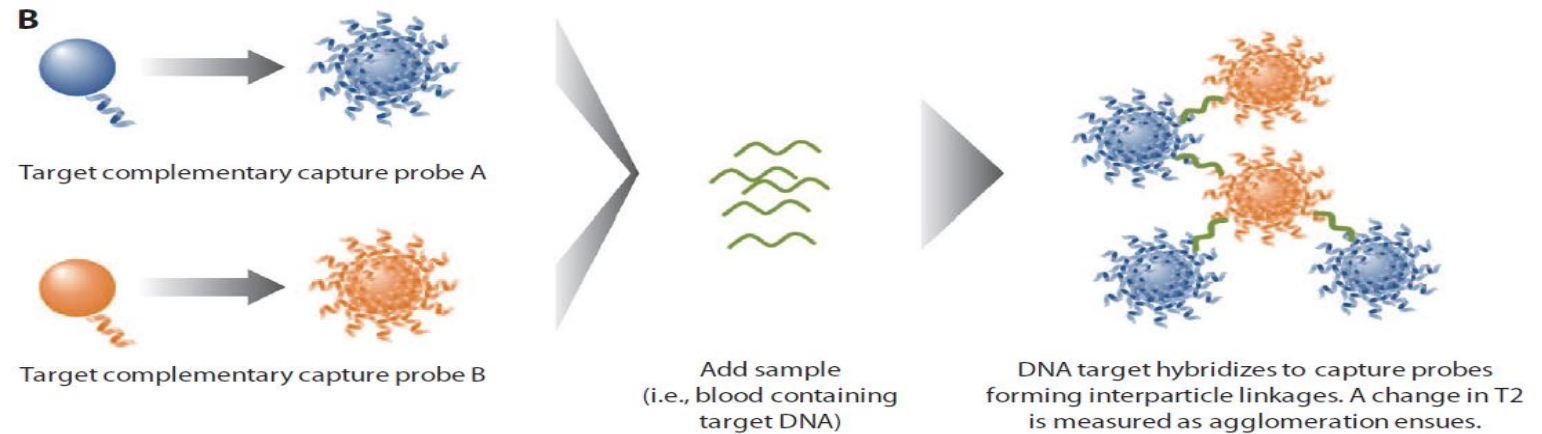
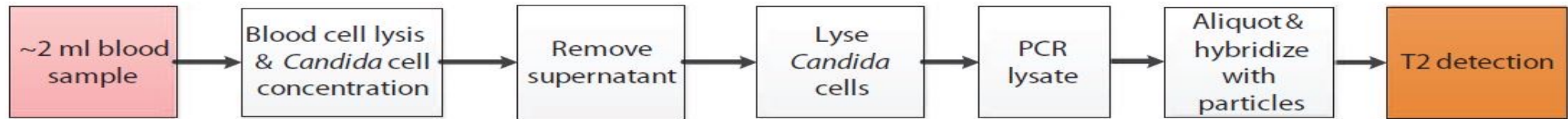
# (1,3)- $\beta$ D- glucans in IC diagnosis

References	Patient type	Number patients/ samples (mean)	IC type (cases)	Cutoff <sup>b</sup>	Sensitivity (%) (95 % CI)	Specificity (%) (95 % CI)	PPV (%) (95 % CI)	NPV (%) (95 % CI)	Proven IC BG <sup>b</sup> (median)
Tissot et al. [38]	Surgical (abdominal) pancreatitis	89/921 [9]	IAC (29)	$\geq 80$	65 (46–82) <sup>a</sup>	78 (63–90) <sup>a</sup>	68 (48–84) <sup>a</sup>	77 (61–88) <sup>a</sup>	223
León et al. [65]	SAC	176/766 (4.3)	C, IAC (31)	$\geq 80$	51.6 (34–69)	86.9 (78–92)	59.3 (40–75)	83.0 (73–89)	259
Del Bono et al. [64]	Surgical	152/152 (1)	C (53)	$\geq 80$	62	98	98.4	57.3	324
Posteraro et al. [63]	Medical/ surgical	95/130 (1.3)	C (13 + 1 M)	$\geq 80$	92.9 (66–99)	93.7 (85–90)	72.2 (46–90)	98.7 (92–99)	500
Mohr et al. [62]	Surgical	57/239 (4)	C [3]	$\geq 80$	100 <sup>a</sup>	59 <sup>a</sup>	NDA	NDA	171

# T2 Candida for the Diagnosis of Candidemia and Invasive *Candida* Infections

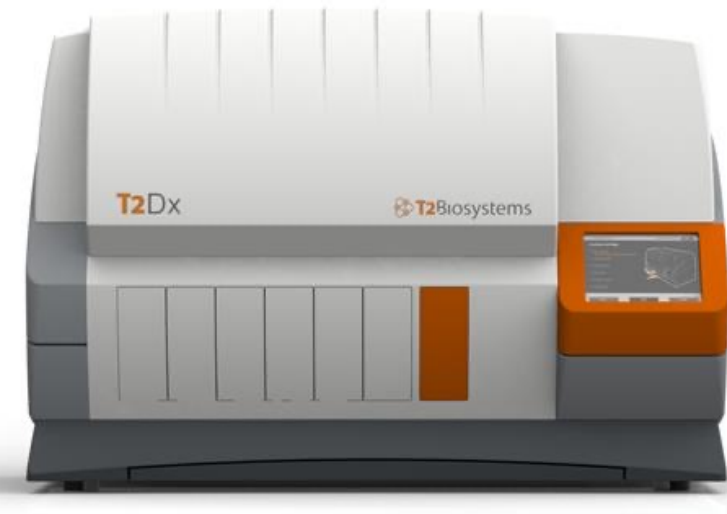


## T2 Candida : workflow



Beyda et al. Diagn Microbiol Infect Dis. 2013; Neely et al. 2013. Sci Transl Med;  
Mylonakis et al. 2015. CID; Bilir et al. 2015. Future Microbiol

# T2 Candida for the Diagnosis of Candidemia and Invasive *Candida* Infections



T2Candida® Panel: US FDA approved and CE marked Nucleic Acid Amplification (NAA) platform for the qualitative detection by magnetic resonance of 5 common *Candida* species :

*C. albicans/C. tropicalis*, *C. parapsilosis*, *C. krusei/C. glabrata* (95% of *Candida* isolates)

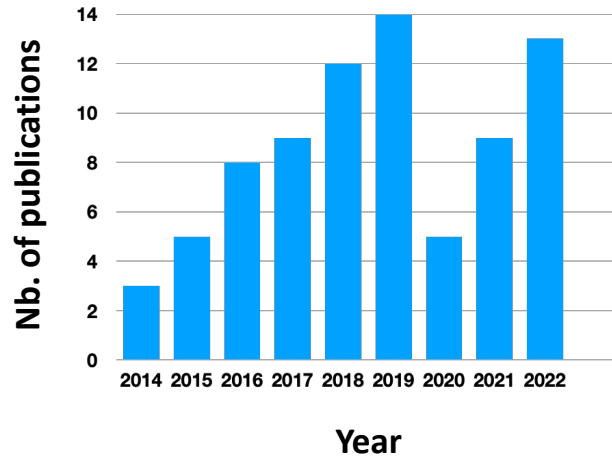
- Fully automated (T2Dx Instrument; T2 Biosystems®), 5 mL Whole blood (EDTA).
- Turn around time for analysis: < 5h
- Limit of detection: 1 - 3 CFU/mL



Neely et al. 2013. *Sci Transl Med*; Beyda et al. *Diagn Microbiol Infect Dis*. 2013; Mylonakis et al. 2015. *CID*; Bilir et al. 2015. *Future Microbiol*

# T2 Candida for the Diagnosis of Candidemia

77 publications since 2014



Clinical Infectious Diseases

MAJOR ARTICLE



## Detecting Infections Rapidly and Easily for Candidemia Trial, Part 2 (DIRECT2): A Prospective, Multicenter Study of the T2Candida Panel

Cornelius J. Clancy,<sup>1</sup> Peter G. Pappas,<sup>2</sup> Jose Vazquez,<sup>3</sup> Marc A. Judson,<sup>4</sup> Dimitrios P. Kontoyiannis,<sup>5</sup> George R. Thompson III,<sup>6</sup> Kevin W. Garey,<sup>7</sup> Annette Reboli,<sup>8</sup> Richard N. Greenberg,<sup>9</sup> Senu Apewokin,<sup>10</sup> G. Marshall Lyon III,<sup>11</sup> Luis Ostrosky-Zeichner,<sup>12</sup> Alan H. B. Wu,<sup>13</sup> Ellis Tobin,<sup>14</sup> M. Hong Nguyen,<sup>1</sup> and Angela M. Caliendo<sup>15</sup>

T2C was sensitive for diagnosing candidemia at the time of positive BC. In patients receiving antifungal therapy, T2C identified bloodstream infections that were missed by companion BCs.

Clancy et al. Clin Infect Dis 2018

MAJOR ARTICLE

## T2 Magnetic Resonance Assay for the Rapid Diagnosis of Candidemia in Whole Blood: A Clinical Trial

Eleftherios Mylonakis,<sup>1</sup> Cornelius J. Clancy,<sup>2</sup> Luis Ostrosky-Zeichner,<sup>3</sup> Kevin W. Garey,<sup>4</sup> George J. Alangaden,<sup>5</sup> Jose A. Vazquez,<sup>6</sup> Jeffrey S. Groeger,<sup>7</sup> Marc A. Judson,<sup>8</sup> Yuka-Marie Vinagre,<sup>9</sup> Stephen O. Heard,<sup>10</sup> Fainareti N. Zervou,<sup>1</sup> Ioannis M. Zacharioudakis,<sup>1</sup> Dimitrios P. Kontoyiannis,<sup>11</sup> and Peter G. Pappas<sup>12</sup>

First prospective multicenter study (Direct 1) : 1801 hospitalized patients who had a BC ordered for routine standard of care; 250 of them were manually supplemented with concentrations from <1 to 100 colony CFUs/mL for 5 different Candida species :

**Sensitivity/Specificity : 91.1%/ 99,4%,**

Sensitivity : 88.17% for subgroup K/G

92.3% for subgroup A/T

94,27% for P

Mylonakis et al.

Clin Infect Dis 2015

J Antimicrob Chemother 2018; 73 Suppl 4: iv27–iv30  
doi:10.1093/jac/dky046

Journal of Antimicrobial Chemotherapy

## Impact of rapid, culture-independent diagnosis of candidaemia and invasive candidiasis in a community health system

M. E. Patch<sup>1\*</sup>, E. Weisz<sup>1</sup>, A. Cubillos<sup>1</sup>, S. J. Estrada<sup>1,2</sup> and M. A. Pfaller<sup>3</sup>

- The implementation of T2Candida improves time to appropriate antifungal therapy for candidaemic patients (34 h for BC versus 6 h for T2Candida, P = 0.0147)

Patch et al. JAC 2018

# T2 Candida for the Diagnosis of Invasive *Candida* Infections

## Performances of T2C in the detection of invasive candidiasis in combination with $\beta$ -glucans and/or mannan

Open Forum Infectious Diseases

### BRIEF REPORT

## Performance of the T2Candida Panel for the Diagnosis of Intra-abdominal Candidiasis

Frederic Lamoth<sup>1,2,a</sup>, Cornelius J. Clancy<sup>3,a</sup>, Frederic Tissot<sup>1</sup>, Kevin Squires<sup>3</sup>, Philippe Eggimann<sup>4</sup>, Ursula Flückiger<sup>5</sup>, Martin Siegemund<sup>6</sup>, Christina Orasch<sup>7</sup>, Stefan Zimmerli<sup>8</sup>, Thierry Calandra<sup>1</sup>, Oscar Marchetti<sup>1,9</sup>, Minh H. Nguyen<sup>3,b</sup>, and Pierre-Yves Bochud<sup>1,b</sup>, on behalf of the Fungal Infection Network of Switzerland (FUNGINOS)

**Table 1. Performance of T2Candida and 1,3-Beta-D-Glucan for Detection of Intra-abdominal Candidiasis**

	T2Candida	1,3-Beta-D-Glucan
Sensitivity,	33.3 (13.3–59.0)	83.3 (58.6–96.4)
Specificity, %	93.3 (77.9–99.2)	66.7 (47.2–82.7)
Positive predictive value, <sup>a</sup> %	71.1 (35.7–91.6)	55.2 (41.6–68.0)
Negative predictive value, <sup>a</sup> %	74.0 (66.9–80.0)	89.0 (73.7–95.9)
Positive likelihood ratio	5.00 (1.13–22.18)	2.50 (1.45–4.32)
Negative likelihood ratio	0.71 (0.51–1.00)	0.25 (0.09–0.72)

Values in brackets are 95% confidence intervals.

Cohort of 48 IAC patients : IAC was **present** in **100%** of cases with **concordant positive** T2Candida/1,3-beta-D-glucan and **absent** in 90% of **concordant negative** results.

Clinical Infectious Diseases

### MAJOR ARTICLE



418 specimen pairs (406 enrolled, 12 spiked) included in analysis

## Multicenter Prospective Study of Biomarkers for Diagnosis of Invasive Candidiasis in Children and Adolescents

Brian T. Fisher,<sup>1,2</sup> Craig L. K. Boge,<sup>1</sup> Rui Xiao,<sup>2</sup> Sydney Shuster,<sup>1</sup> Dawn Chin-Quee,<sup>3</sup> John Allen IV,<sup>3</sup> Shareef Shaheen,<sup>3</sup> Randall Hayden,<sup>4</sup> Sri Suganda,<sup>4</sup> Theoklis E. Zaoutis,<sup>1,2</sup> Yeh-Chung Chang,<sup>3</sup> Dwight E. Yin,<sup>5</sup> Anna R. Huppler,<sup>6</sup> Lara Danziger-Isakov,<sup>7</sup> William J. Muller,<sup>8</sup> Emmanuel Roilides,<sup>9</sup> José Romero,<sup>10</sup> Paul K. Sue,<sup>11</sup> David Berman,<sup>12</sup> Rachel L. Wattier,<sup>13</sup> Natasha Halasa,<sup>14</sup> Alice Pong,<sup>15</sup> Gabriela Maron,<sup>16</sup> Pere Soler-Palacin,<sup>17</sup> Susan C. Hutto,<sup>18</sup> Blanca E. Gonzalez,<sup>19</sup> Christine M. Salvatore,<sup>20</sup> Sujatha Rajan,<sup>21</sup> Michael Green,<sup>22</sup> Elizabeth Doby Knackstedt,<sup>23</sup> Sarmistha B. Hauger,<sup>24</sup> and William J. Steinbach<sup>3</sup>

**Table 7. Sensitivity, Specificity, Positive Predictive Value, and Negative Predictive Value for the T2Candida and Platelia *Candida* Antigen Plus Results Used in Combination**

Test	Sensitivity	Specificity	Positive Predictive Value	Negative Predictive Value	Specimens Exceeding Cutoff
	% (95% CI)	% (95% CI)	% (95% CI)	% (95% CI)	n
T2Candida only	77.3 (54.6, 92.2)	97.2 (95.1, 98.6)	60.7 (40.6, 78.5)	98.7 (97.0, 99.6)	28
Platelia <i>Candida</i> Antigen Plus only	40.9 (20.7, 63.7)	97.2 (95.1, 98.6)	45.0 (23.1, 68.5)	96.7 (94.5, 98.3)	20
At least one test positive	86.4 (65.1, 97.1)	94.7 (92.0, 96.7)	47.5 (31.5, 63.9)	99.2 (97.7, 99.8)	40
Both tests positive	31.8 (13.9, 54.9)	99.8 (98.6, 100.0)	87.5 (47.4, 99.7)	96.3 (94.0, 97.9)	8

A total of 418 specimen pairs (406 enrolled, 12 spiked) were included in analysis : **T2Candida alone or in combination with mannan antigen** may be beneficial for rapid detection of *Candida* species in children with concern for IC.

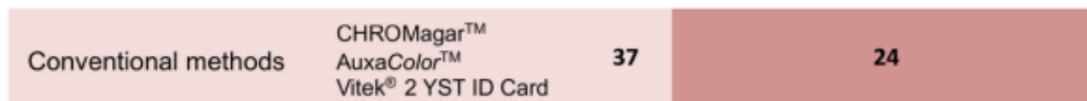
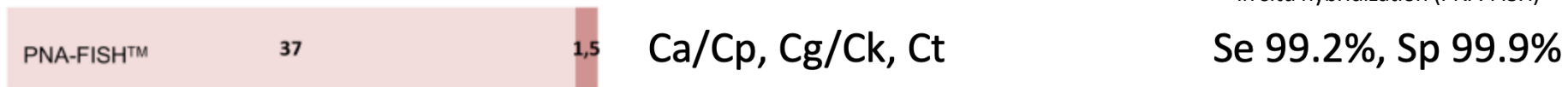
# Time to yeast identification

Ibáñez-Martínez et al. Rev Esp Quimioter 2017

## WHOLE BLOOD METHODS



## BLOOD CULTURE BASED METHODS



Conventional methods + MALDI-TOF = 24-30 depending on Candida species

0 12 24 36 48 60 72  
Time (hours)

Figure 2 | Graphic representation of time (hours) to yeast identification depending on employed technique.



## **Diagnostic biologique des aspergilloses invasives**

# Diagnostic biologique des aspergilloses invasives

- Mycologie conventionnelle
- Antigène galactomannane dans le sérum et le LBA (+LCR)
- Lateral Flow Device? (situation d'urgence)
- $\beta$ -(1,3)-D-glucanes

# Aspergillose invasive- Mycologie conventionnelle

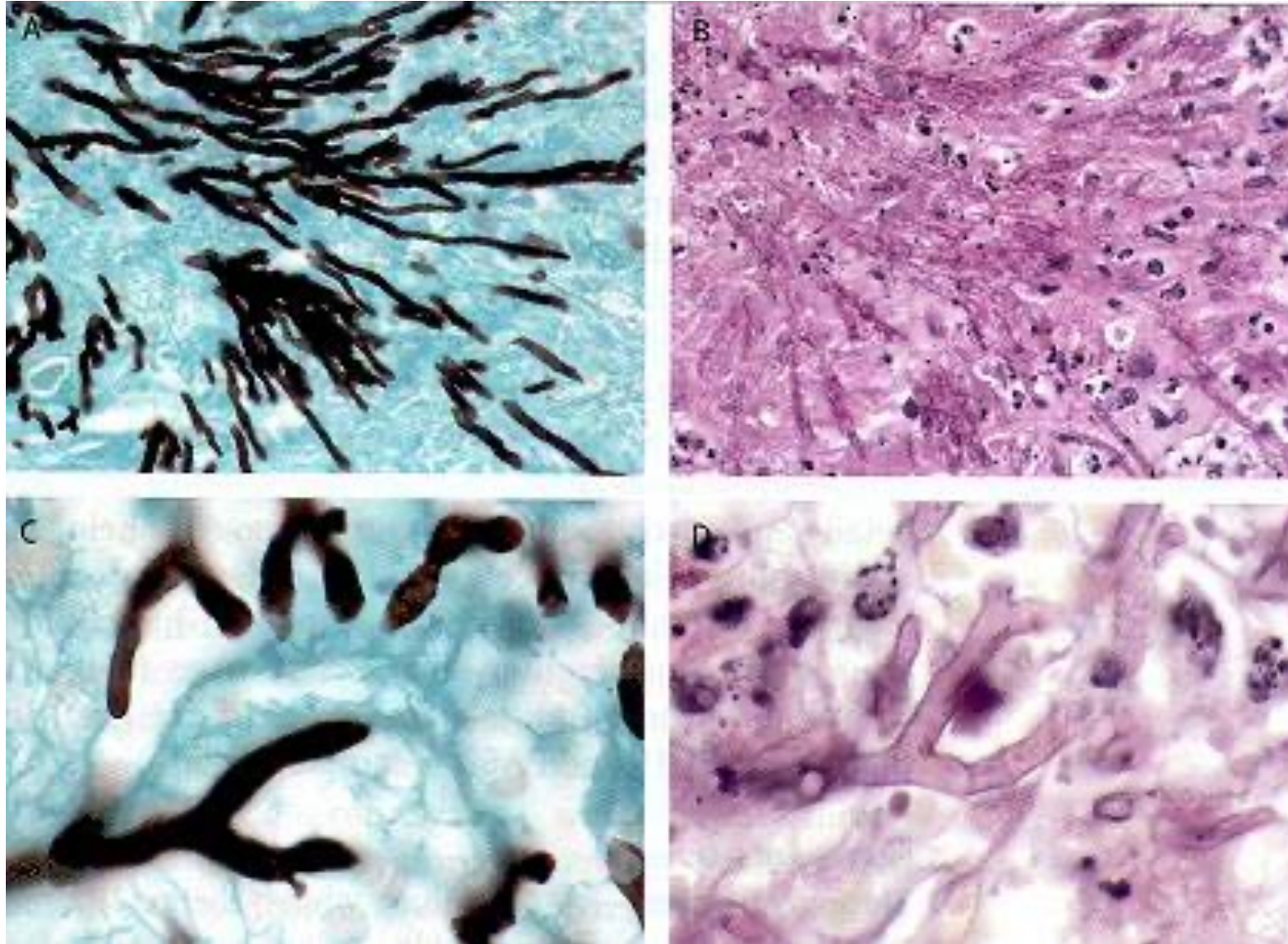
- **Diagnostic conventionnel**

**Examen direct des  
tissus**

**prélèvements endobronchiques (crachat,  
aspirations trachéales, LBA)**

	<u>Crachat/LBA</u>	<u>tissus</u>
Etat frais	préparation ± KOH	
Colorations de routine	Gram	HE
Colorations spécifiques	GMS, PAS, BT	GMS, PAS
Agents fluorescents	Calcofluor white	
Colorant fluorescent	Uvitex 2B Blankophor	

# Aspergillose invasive- Diagnostic Conventionnel



In: Hope et al., *Lancet Infectious Diseases* 5: 609, 2005

# Aspergillose invasive- Diagnostic Conventionnel

## DIAGNOSTIC HISTOLOGIQUE

Améliorer la détection des filaments par immunohistochimie

Anticorps monoclonaux

- WF-AF-1 (Dako) <sup>1)</sup>
- EB-A1 <sup>2)</sup>

Sensibilité pour AI prouvée par la culture : 89-94%

Résultats spécifiques de genre ou d'espèce

1) Choi JK et al., *Am J Clin Pathol* 121: 18, 2004

2) Pierard GE et al., *Am J Clin Pathol* 96: 373, 1991

Verweij PE et al., *Am J Clin Pathol* 49: 798, 1996

# Galactommanane sérique

Polysaccharide libéré par les champignons du genre *Aspergillus* au cours de leur croissance  
Méthode de détection standardisée : kit ELISA Platelia Aspergillus Ag (Bio-Rad®)



54 études, 5 660 patients, 586 avec une forme prouvée ou probable (EORTC/MSG)

Patients neutropéniques

→ DOI à 0.5 (**Se / Sp**) : **78% / 85%**

(Leeflang *et al.*, Cochrane Database Syst Rev, 2015)

1 109 patients admis en réanimation

26 formes prouvées

Remarque : 10 patients neutropéniques

→ DOI à 0.5 : **Se de 42%**

(Meersseman *et al.*, Am J Respir Crit Care Med, 2008)

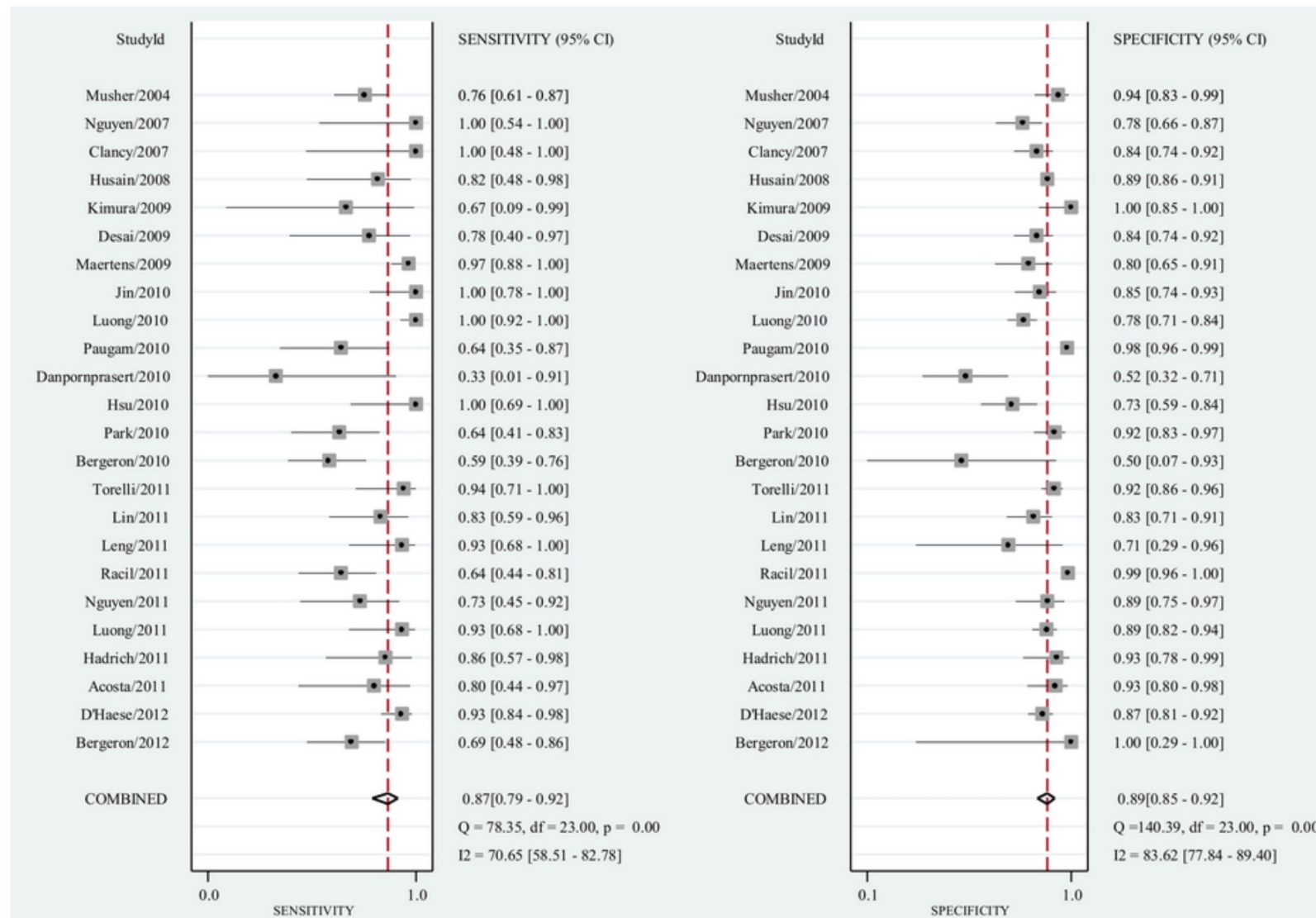
Analyse de 126 cas d'AI chez des transplantés d'organes solides (poumon, cœur, foie, rein)

Incidence de 6,5%, 2,9%, 1,8 et 0,6% respectivement

→ DOI à 0.5 : **Se de 50,6%**

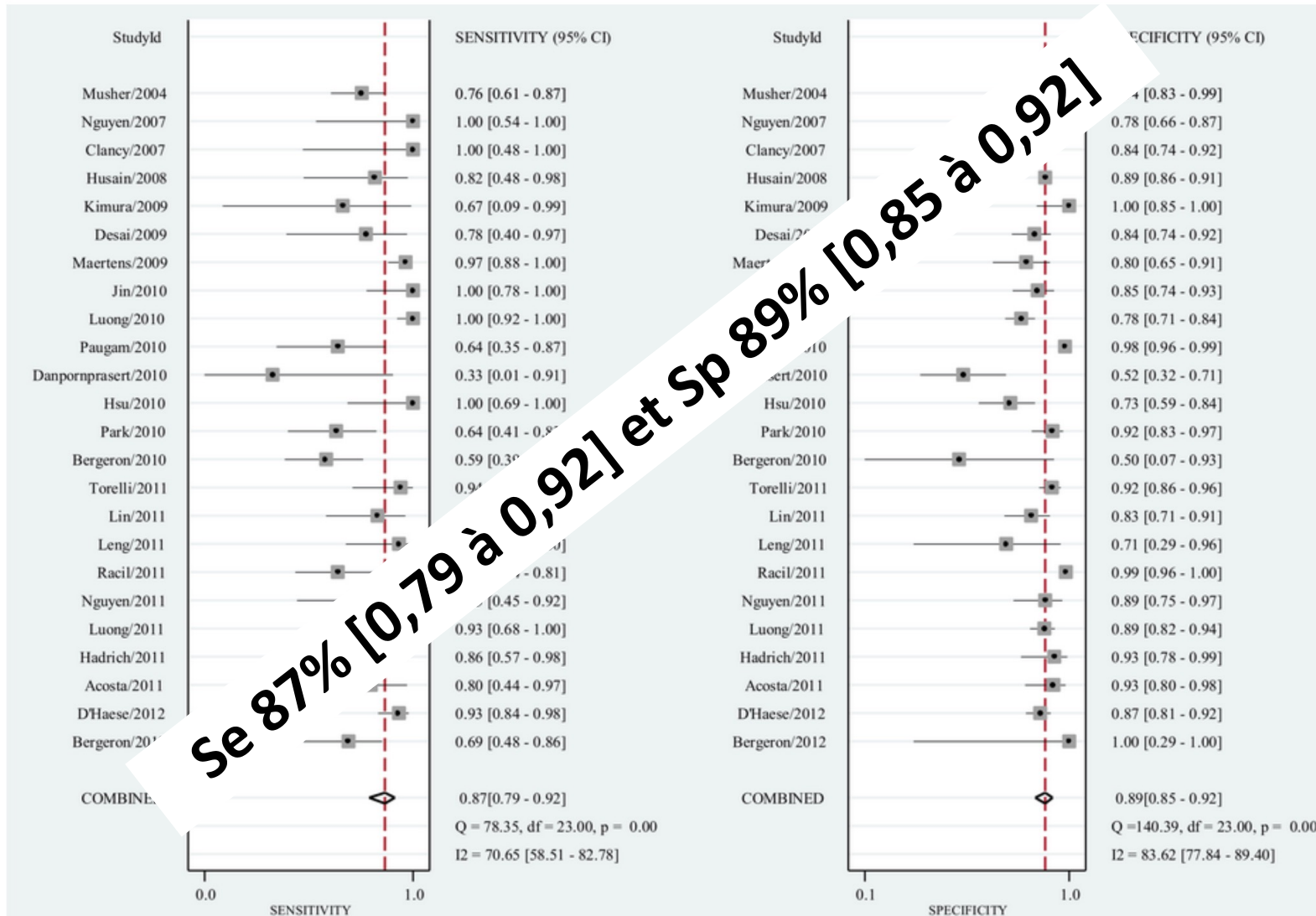
(Gioia *et al.*, Mycoses, 2021)

# Détection du galactommanane dans le LBA



(Zou *et al.*, Plos One, 2012)

# Détection du galactommanane dans le LBA



(Zou et al., Plos One, 2012)



## Apport de la PCR au diagnostic des API

→ Entrée de la PCR *Aspergillus* dans les critères EORTC/MSG du diagnostic des API (Donnelly *et al.*, Clin Infect Dis., 2020)

PCR positive à deux reprises dans le plasma, le sérum ou le sang total

PCR positive dans le LBA à deux reprises ou en dupliqua

Association d'une PCR sérique/plasma/sang total positive et d'une PCR dans le LBA

# Apport de la PCR au diagnostic des API

- PCR: manque de standardisation
  - ADN circulant: Se 84% Sp 76%
  - LBA: Se 77% Sp 94%

Méta-analyse Cochrane : 18 études - 2000-2013.

– 19 cohortes

- Patients onco-hémato (adultes-enfants)
- Haut risque IA → Prévalence moy de IA (prouvée ou probable ) : 12% (2.5 à 30.8 %).

– Différentes techniques de qPCR (PCR maison).

- Cible : *A. fumigatus*

– Valeurs intrinsèques

Critère diagnostique	Sensibilité moy (IC 95%)	Spécificité moy (IC 95%)
1 PCR pos	80,5 (73 - 86.3)	78.5 (67.8 - 86.4)
2 PCR pos	58 (36.5 -76.8)	96 (89.6 - 98.6)

# Apport de la PCR au diagnostic des API

## Performances variables selon les populations étudiées

- Patients d'hématologie à haut risque

25 études, 2 595 patients inclus

Pour un répliquat **Se / Sp : 84% / 76%**

Pour deux PCR consécutives positives **Se / Sp : 64% / 95%**

(Arvanitis *et al.*, JCM, 2014)

- HM, HSCT, SOT

34 études

Pour un répliquat **Se / Sp : 79,2% / 79,6%**

Pour deux PCR consécutives positives **Se / Sp : 59,6% / 95,1%**

Très bonne VPN (95%)

(Cruciani *et al.*, Cochrane Library, 2019)

- Analyse en sous-groupe réalisée sur la cohorte précédente pour déterminer l'impact de la chimioprophylaxie sur les performances de la PCR.

→ Pas de perte de Se, par contre perte de Sp (perte de 26% si l'on considère une seule PCR positive, perte de 12% si l'on considère deux ou plus résultats de PCR)

(Cruciani *et al.*, J Antimicrob Chemother, 2021)

# Apport de la PCR au diagnostic des API

## Tests commerciaux

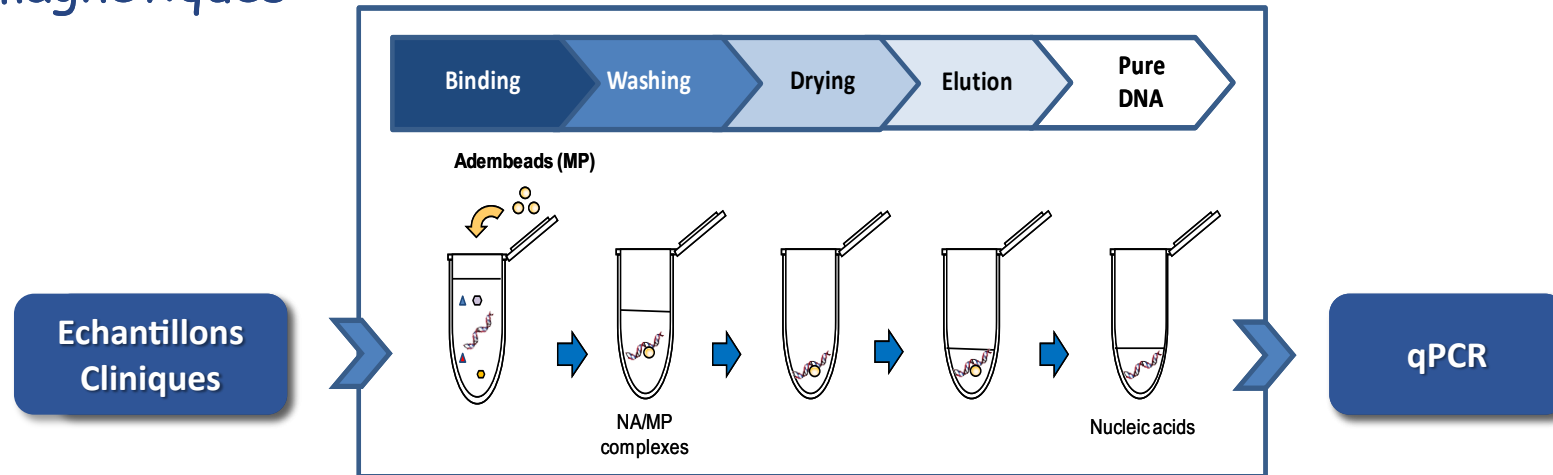
- MycoGENIE (Ademtech), AsperGenius (PathoNostics), Fungiplex (Renishaw).
- Sang, LBA, +/-biopsies
- AsperGenius (PathoNostics):
  - 2 multiplex
  - 5 espèces aspergillaires + mutation TR34, L98H, T289A, et Y121F (CYP51A)
  - Se 84.2%, Sp 91.4%, VPP 76.2%, and VPN 94.6%

Nécessité d'évaluation prospective multicentrique pour préciser les performances diagnostiques dans différentes populations de patients

# MycoGENIE *Aspergillus* : Détection d'ADN *A. fumigatus* dans le sérum

Détection d'ADN d'*Aspergillus fumigatus* et de la mutation TR34/L98H

## ❑ Purification d'Acides Nucléiques à l'aide de billes magnétiques



- ✓ Automatisé
- ✓ Pas de centrifugation
- ✓ ADN de grande qualité
  - Pas de dégradation
  - Pas d'inhibiteurs PCR
  - Grande concentration

- Plaques pré-remplies
- Prêtes à l'emploi
- 12 extractions / run
- Durée 35min
- Volume d'élution 50µl



## Performances du test MycoGENIE pour le diagnostic des aspergilloses invasives

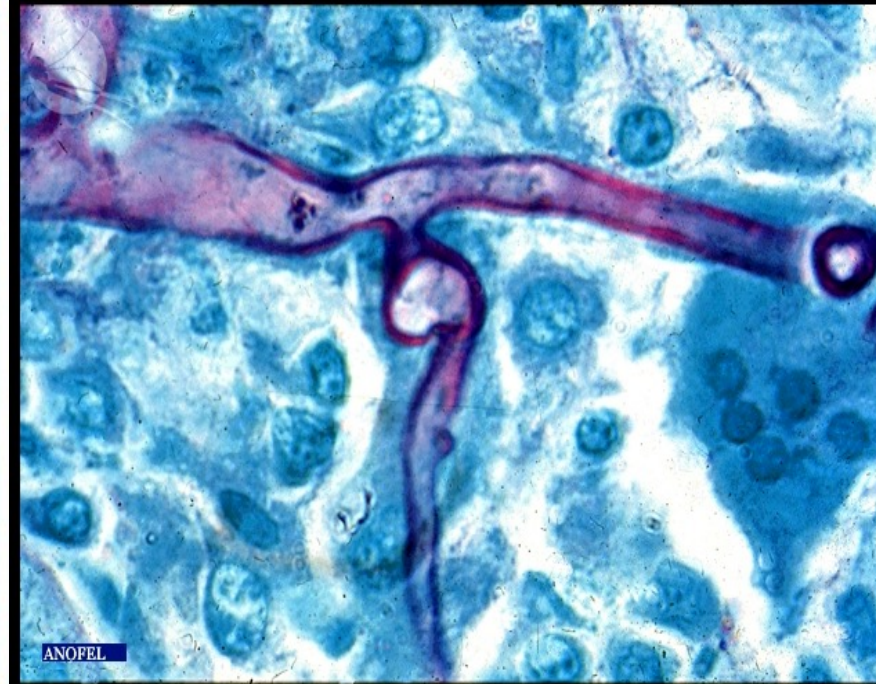
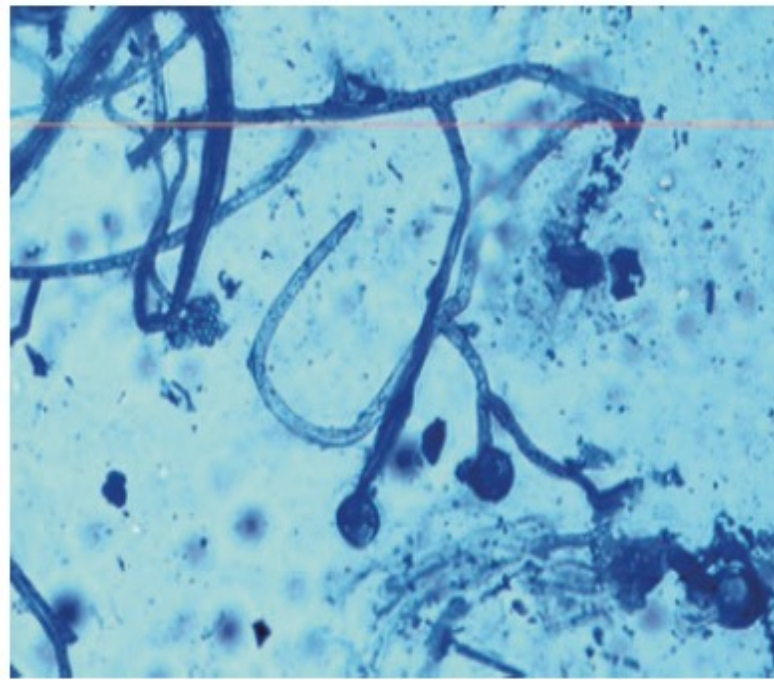
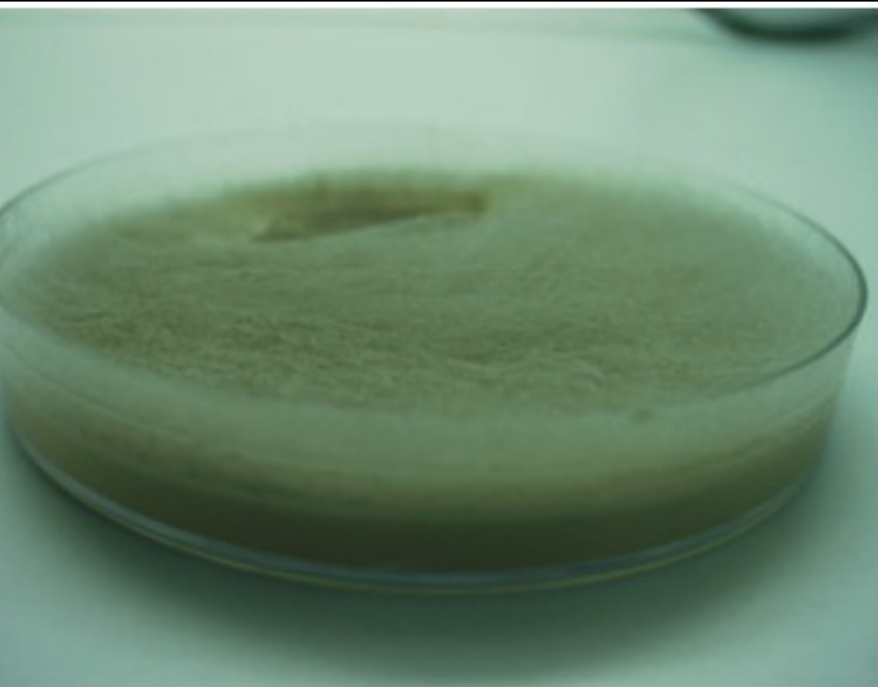
- Dannaoui *et al.*, JCM, 2017 :
  - Respiratoires n=88 / 62 patients : Se 92,9%; Sp : 90,1%
  - Sérums n= 69 / 16 patients AI prouvée-probable et 13 patients non AI
    - Se 100%; Sp 84,6
  - LOD : Aspergillus : 1 copie TR34/L98H : 6 copies
- Denis *et al.*, J.Mol. Diagn., 2018
  - LBA n=73 / 31 AI probables avec Se 100%; Sp : 71%

# EORTC/MSGERC IPA case definitions

Criteria	Clinical	Radiological	Mycological
<b>Proven IPA</b>	-	-	<b>Lung biopsy, at least 1:</b> <ul style="list-style-type: none"> <li>Histo/cytopathologic or direct microscopic examination (hyphae + tissue damage)</li> <li>Positive culture from tissue</li> </ul>
<b>EORTC/MSGERC</b> Donnelly JP, CID 2020	<b>Host factors:</b> <ul style="list-style-type: none"> <li>Neutropenia</li> <li>Malignant hemopathy</li> <li>Transplant</li> <li>Prolonged corticosteroids &gt;0.3mg/kg &gt;3weeks/2months</li> <li>Immunosuppressive drugs...</li> </ul>	<b>At least 1 CT pattern:</b> <ul style="list-style-type: none"> <li>Dense, well-circumscribed lesion (<math>\pm</math>halo)</li> <li>Air crescent sign</li> <li>Cavity</li> <li>Consolidation</li> </ul>	<b>At least 1:</b> <ul style="list-style-type: none"> <li>Positive direct microscopy or culture of a respiratory sample (sputum, tracheal aspirate, BAL)</li> <li>BAL GM <math>\geq 1</math></li> <li>serum GM <math>\geq 1</math></li> <li>BAL GM <math>\geq 0.8</math> <u>and</u> serum GM <math>\geq 0.7</math></li> <li><u>2x</u> positive <i>Aspergillus</i> PCR (serum or BAL)</li> </ul>
<b>Possible IPA</b>	<i>idem</i>	<i>idem</i>	-

# Diagnostic des mucormycoses invasives

- Mycologie conventionnelle
- Histologie +++
- Pas de test de détection de polysaccharides circulants





# Diagnostic des mucormycoses invasives

## Détection d'ADN circulant par PCR pour le diagnostic des zygomycoses

JOURNAL OF CLINICAL MICROBIOLOGY, Nov. 2008, p. 3690–3702  
0095-1137/08/\$08.00+0 doi:10.1128/JCM.00917-08  
Copyright © 2008, American Society for Microbiology. All Rights Reserved.

Vol. 46, No. 11



### Detection of a Molecular Biomarker for Zygomycetes by Quantitative PCR Assays of Plasma, Bronchoalveolar Lavage, and Lung Tissue in a Rabbit Model of Experimental Pulmonary Zygomycosis<sup>∇</sup>

Miki Kasai,<sup>1†</sup> Susan M. Harrington,<sup>2†‡</sup> Andrea Francesconi,<sup>1</sup> Vidmantas Petraitis,<sup>1,3</sup> Ruta Petraitiene,<sup>1,3</sup> Mara G. Beveridge,<sup>1</sup> Tena Knudsen,<sup>1</sup> Jeffery Milanovich,<sup>1</sup> Margaret P. Cotton,<sup>1</sup> Johanna Hughes,<sup>1</sup> Robert L. Schaufele,<sup>1</sup> Tin Sein,<sup>1</sup> John Bacher,<sup>4</sup> Patrick R. Murray,<sup>2</sup> Dimitrios P. Kontoyiannis,<sup>5</sup> and Thomas J. Walsh<sup>1\*</sup>

### Quantitative Polymerase Chain Reaction Detection of Circulating DNA in Serum for Early Diagnosis of Mucormycosis in Immunocompromised Patients

Laurence Millon,<sup>1,2</sup> Fabrice Larosa,<sup>3</sup> Quentin Lepiller,<sup>2,4</sup> Faezeh Legrand,<sup>3</sup> Steffi Rocchi,<sup>1</sup> Etienne Daguindau,<sup>3</sup> Emeline Scherer,<sup>1,2</sup> Anne-Pauline Bellanger,<sup>1,2</sup> Joel Leroy,<sup>5</sup> and Frederic Grenouillet<sup>1,2</sup>

# Diagnostic des mucormycoses invasives

## La détection d'ADN peut précéder de plusieurs jours la positivité de l'histologie

**Table 1 Clinical, Histological, and Mycological Findings, Antifungal Treatment, Results of Quantitative Polymerase Chain Reaction Assays, and Outcomes for 10 Patients With Proven Mucormycosis**

Patient No.	Sex <sup>a</sup> / Age	Underlying Disease	Localization of Infection, Date <sup>b</sup> of First Clinical Symptoms/First Radiological Signs	Date <sup>b</sup> of Fever Onset	Positive Histology (Day 0)/Molecular Diagnosis	Positive Mycological Cultures (Day)/ Identification	Antifungal Therapy (Day of Initiation of L-AmB, POS)	Outcome at Day 90	qPCR Result Cq (Day)	Days From 1st Positive PCR to 1st Clinical Signs <sup>c</sup>	Days From 1st Positive PCR to Positive Histology <sup>d</sup>
1	F/65	Undernutrition, alcoholism	Disseminated D -1/not done	D -5	Skin/not done	Skin, urine (D0)/ <i>L. corymbifera</i>	FLU	Death (D1)	Acory Neg (D -8, D -1)	/	/
2	M/37	HL	Disseminated D -16/not done	D -28	Lung, liver/not done	Lung, liver (D0)/ <i>L. corymbifera</i>	L-AmB (D -1)	Death (D0)	Acory 28 (D -3), Neg (D -10)	+13	-3
3	M/51	ALL	Rhinocerebral D -1/D -1	D -9	Sinus/ <i>Lichtheimia corymbifera</i>	Sinus (D0)/ <i>L. corymbifera</i>	L-AmB (D0) POS (D0)	Death (D3)	Acory 36 (D -8), 32 (D -5), 30 (D -1), 25 (D0), 27 (D2), Neg (D -18, D -15, D -12)	-7	-8
4	F/48	Polytrauma	Cutaneous D -5/not done	D0	Skin/ <i>Lichtheimia</i> sp	Skin (D0)/negative	FLU, CAS	Death (D2)	Acory 27 (D0), Neg (D -2, D -9)	+5	0
5	M/59	NHL, diabetes mellitus	Disseminated D -5/D -22	D -14	Kidney/ <i>Lichtheimia</i> sp	Cerebrospinal fluid (D -10)/ <i>Lichtheimia ramosa</i>	FLU, CAS, L-AmB (D0)	Alive	Acory 32 (D -23) 38 (D -8), Neg (D -1, D5)	-18	-23
6	F/41	PNH, SAA	Disseminated D -30/D -4	D -17	Sinus/ <i>Rhizopus</i> sp	Sinus (D0)/ <i>R. oryzae</i>	CAS, POS (D -11) L-AmB (D0) deferasirox (D0)	Death (D15)	Muc1 36 (D -10), 35 (D -1), 38 (D2), 36 (D6), 38 (D9), 34 (D13)	+20	-10
7	F/13	AML	Pulmonary <sup>e</sup> D -62/D -61	D -65	Lung/ <i>Rhizopus oryzae</i>	Not done	VOR, L-AmB (D -60)	Alive	Muc1 35 (D -68), 40 (D -62), 36 (D -55), 38 (D -21), 36 (D10), Neg (D -75, D29)	-6	-68
8	M/57	MDS, allo-HSCT	Rhino-cerebral D -18/D -11	D0	Sinus/not done	Sinus (D0)/ <i>R. pusillus</i>	ITR (D -96), L-AmB (D0) POS (D0)	Alive	Rmuc 39 (D -18), 39 (D -1), Neg (D -11, D -8, D9, D13)	0	-18
9	F/55	Renal transplant	Disseminated D -24/D -11	D -30	Lung, heart/ <i>Rhizomucor pusillus</i>	Lung, heart (D0)/ <i>R. pusillus</i>	CAS	Death (D0)	Rmuc 29 (D -4), 22 (D -3), Neg (D -24, D -19)	+20	-4
10	M/60	AML	Disseminated D -50/D -47	D -50	Liver, spleen/ <i>Rhizomucor</i> sp	Liver, spleen (D0)/negative	VOR, L-AmB (D -44) POS (D -36)	Alive	Rmuc 41 (D -49), 28 (D -46), Neg (D -52, D -42, D -39, D -36, D -18, D -13, D -1)	+1	-49

# Diagnostic des mucormycoses invasives

## Early diagnosis and monitoring of mucormycosis by detection of circulating DNA in serum: retrospective analysis of 44 cases collected through the French Surveillance Network of Invasive Fungal Infections (RESSIF)

L. Millon <sup>1, 2, \*</sup>, R. Herbrecht <sup>3</sup>, F. Grenouillet <sup>1, 2</sup>, F. Morio <sup>4, 5</sup>, A. Alanio <sup>6, 7, 8, 9</sup>,  
V. Letscher-Bru <sup>10, 11</sup>, S. Cassaing <sup>12</sup>, T. Chouaki <sup>13, 14</sup>, C. Kauffmann-Lacroix <sup>15</sup>, P. Poirier <sup>16</sup>,  
D. Toubas <sup>17, 18</sup>, O. Augereau <sup>19</sup>, S. Rocchi <sup>2</sup>, D. Garcia-Hermoso <sup>7, 8</sup>, S. Bretagne <sup>6, 7, 8, 9</sup>,  
French Mycosis Study Group

**Effectif** : 19 mucormycoes probables, 25 confirmées (réseau RESSIF)

**Sensibilité** : 81% (au moins un sérum + par patient)

**Précocité du diagnostic** : PCR positive en moyenne 9 jours ( 0-28 jrs) avant la positivité des examens mycologiques; au moins 2 jrs ( 0-24 jrs) avant l'imagerie.

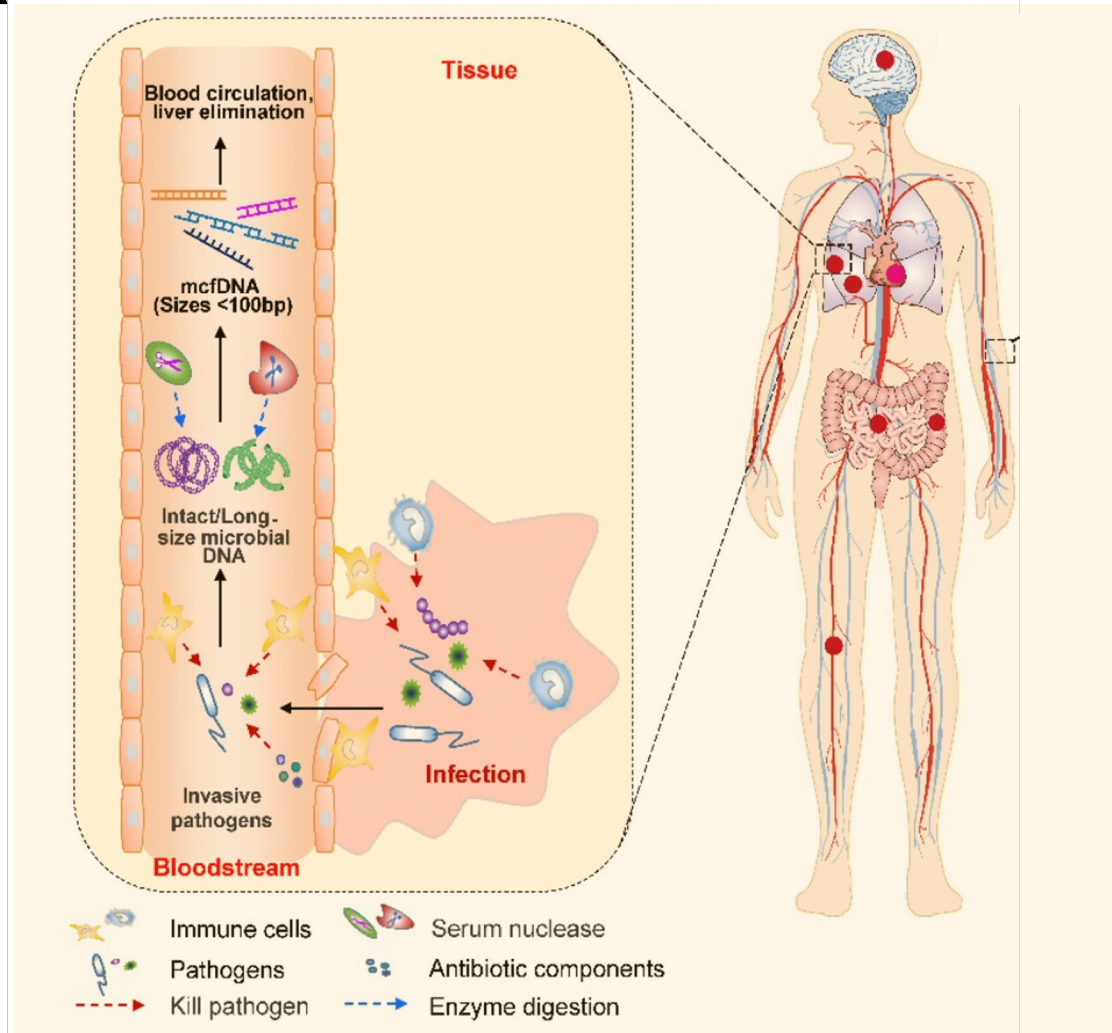
**Valeur pronostique** :

Le taux de survie à 84 jours était significativement plus élevé chez les patients ayant une PCR+ qui se négative sous traitement que ceux pour lesquels la PCR reste constamment positive (48% and 4%, respectivement; p <10<sup>-6</sup>).

## Cell free DNA

In blood and other body fluids

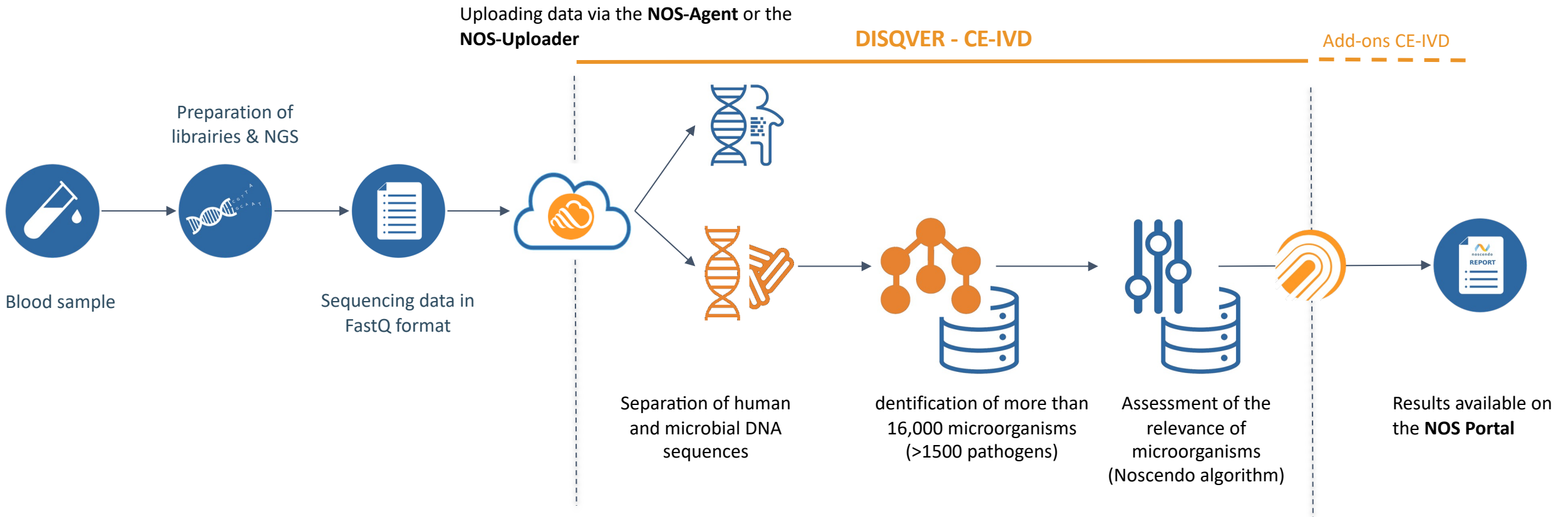
- Urine
- CSF
- Ascites
- Sputum
- BAL
- Synovial fluid
- Other....



Pathogen genetic material is present in the cfDNA fractions of the different body fluids

- Translocation from other compartments

# Methodology



# Estimation of relevance

name	taxonomy_id	taxonomy_lvl	assigned_reads	relevance
<i>Achromobacter</i> sp. AONIH1	1758194	S	2	0.01
<i>Achromobacter xylosoxidans</i>	85698	S	6	0.02
<i>Cloacibacterium normanense</i>	237258	S	1	0.07
<i>Cutibacterium acnes</i>	1747	S	5	0.05
<i>Delftia tsuruhatensis</i>	180282	S	7	0.06
<b><i>Escherichia coli</i></b>	562	S	<b>11</b>	<b>16</b>
<b><i>Homo sapiens</i></b>	9606	S	35,362,170	<b>NA</b>
<b><i>Human gammaherpesvirus 4</i></b>	10376	S	<b>8</b>	<b>16</b>
<i>Microbacterium</i> sp. No. 7	1714373	S	2	0.97
<i>Moraxella osloensis</i>	34062	S	1	0.00
<b><i>Plasmodium ovale</i></b>	36330	S	<b>22</b>	<b>0.03</b>
<b><i>Rhizopus delemar</i></b>	936053	S	<b>47</b>	<b>0.99</b>
<i>Stenotrophomonas maltophilia</i>	40324	S	3	0.02
<b><i>Streptococcus pneumoniae</i></b>	1313	S	<b>1919</b>	<b>16</b>
<i>Wuchereria bancrofti</i>	6293	S	7	0.04

# Available tools for the diagnosis of IFI

Dx Tools	Invasive Candidiasis	Cryptococcosis	Invasive Aspergillosis	Mucormycosis	Fusariosis	Histoplasmosis	Pneumocystosis
Direct examination/ Histology	Morphological aspects of IFI, Poor sensitivity						
Specificities		Capsule Indian Ink, other type of staining					Specific aspects IF
Culture Based Methods	Poor sensitivity						
Antigen	Mannan Spécific Transient Poor sensitivity Serum, (CSF) Diagnosis Deep-seated localisation	Glucuronoxylomannan Sensitivity +++ Sppecificity +++ Serum, CSF (BAL, urine) Diagnosis Prognosis	Galactomannan Sensitivity host- and clinical entity dependant Specificity Cross reaction Other fungi Serum, BAL, CSF (bronchial aspiration) Screening, Diagnosis, Prognosis		Some species cross react with GM	Specific Ag Not available in Fr Cross react with GM	
$\beta$ - D1,3 Glucans	Species dependant		Disappointing		Sensitivity 50%	Scare studies	NPV Long-time persistence
PCR	T2MR (5 species) Blood (CSF/AH) Necker/Marseille (few species)	In house Method	<i>A. fumigatus</i> /spp All matrices Resistance, included in EORTC/MSGERC IPA case definitions	(4 species in Lille)	In house Method	In house Method	Interpretation threshold, diffirenciate infection/colonization
Panfungal Sequencing	Moderate sensitivity if direct examination is positive !						
NGS/WGS	Sensistivity, quantification and interpretation ?						

# Take home messages !

- Diagnostic des mycoses superficielles aisé, rapide
- Délais de rendu des résultats réduits de 24-48 h par rapport aux critères phénotypiques
- Mycologie conventionnelle utile pour l'isolement de l'agent pathogène, réalisation des tests de sensibilité aux antifongiques
- Diagnostic des mycoses invasive est difficile (Mycologie conventionnelle + histologie insuffisants) :
  - Candidoses invasives : Recours aux biomarqueurs (mannane, b-glucanes), T2MR pour améliorer la sensibilité des hémocultures
  - Aspergilloses invasives : Recours au galactomannane sérique, LBA, PCR in situ et PCR dans le sang
  - Mucormycoses : recours à la PCR (améliorer la sensibilité et la précocité du diagnostic)